

The use of organic and inorganic amendments in the phytoremediation of contaminated soils and their effect on biodiversity

Rafael Clemente, CEBAS-CSIC, Murcia (Spain)

School of Soil Biodiversity and Bioindication

Biodiversity and bioindicators in monitoring and management of contaminated soils



UNIVERSITAT DE MURCIA
UNIVERSITY OF MURCIA



DIPARTIMENTO DI
AGRARIA

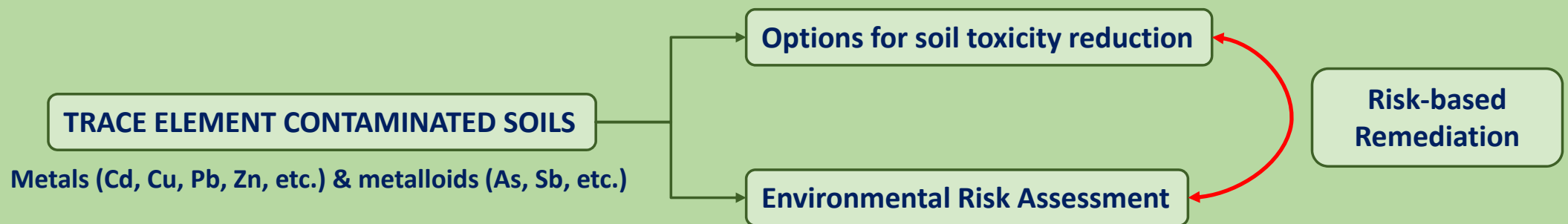


GOBIERNO
DE ESPAÑA

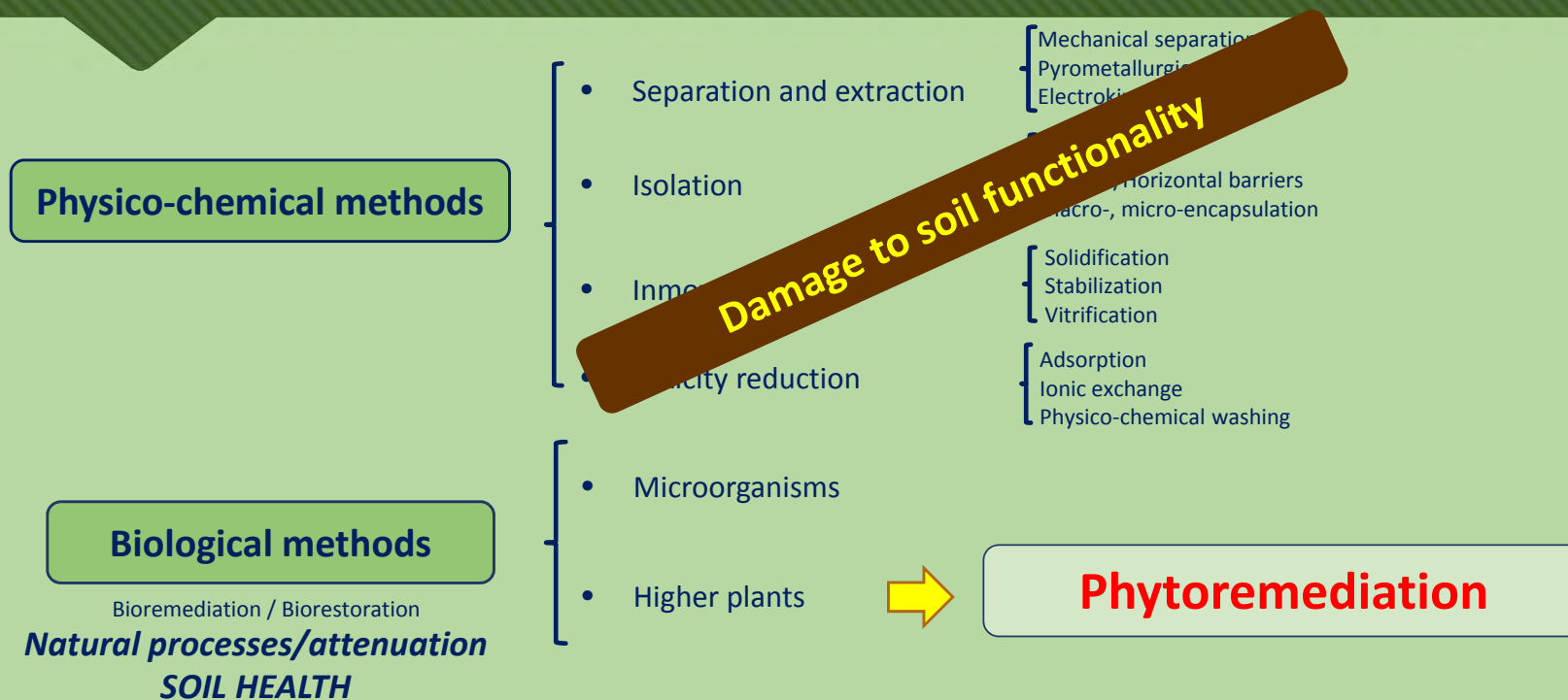
MINISTERIO
DE CIENCIA, INNOVACIÓN
Y UNIVERSIDADES



Trace elements - Contamination - Phytoremediation

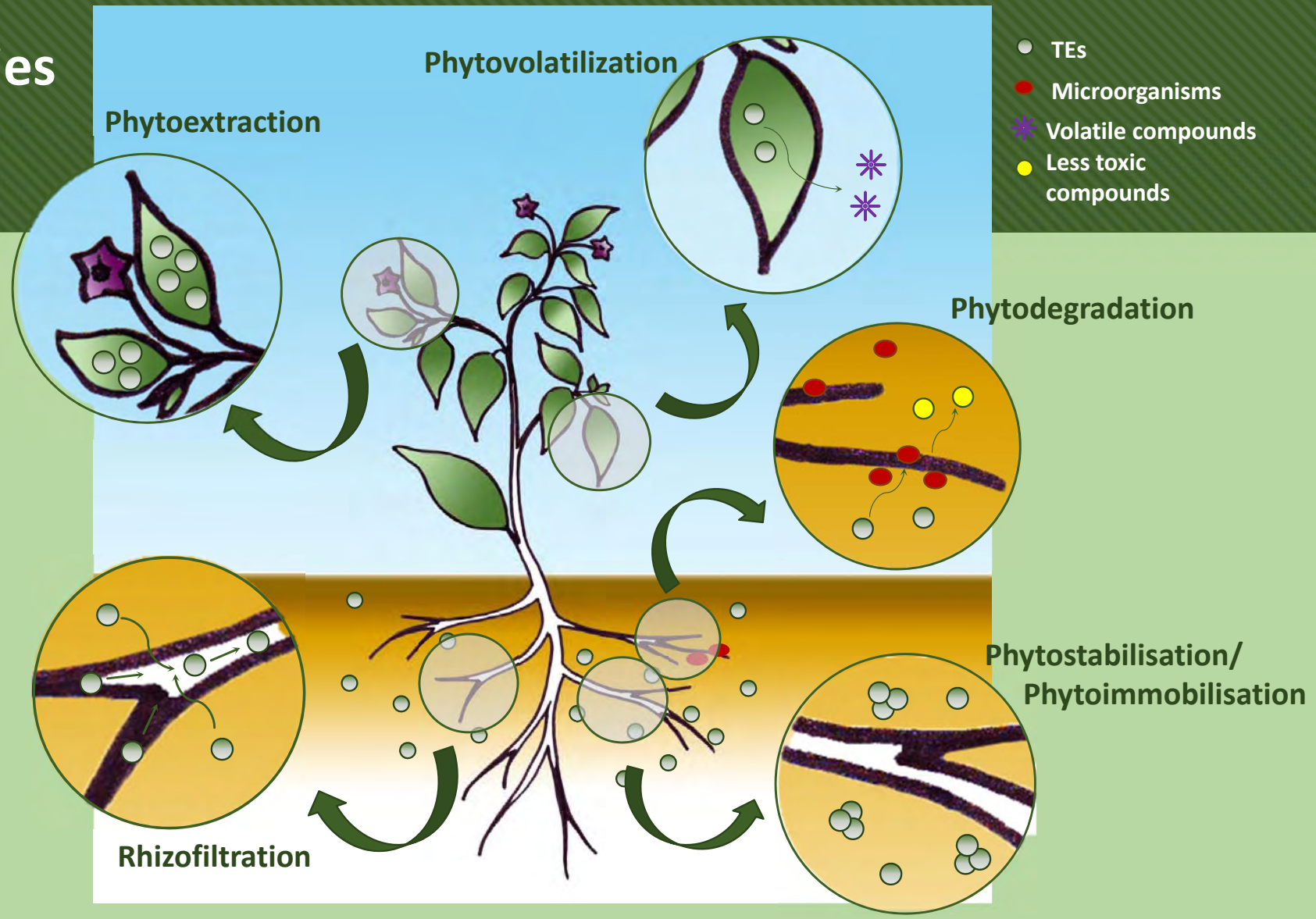


Remediation methods for contaminated soils



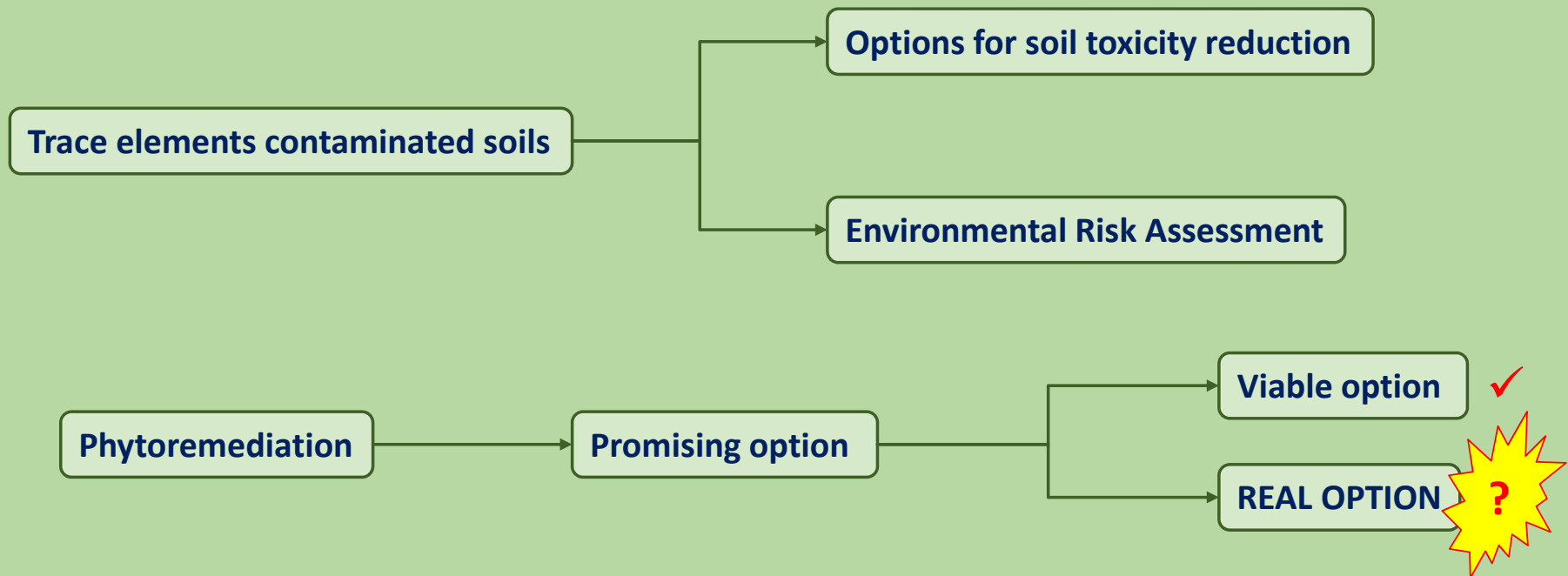
“techniques based on the use of plant species (and their associated microorganisms), **amendments** and agronomical techniques to eliminate, retain or diminish the toxicity of the contaminants” (Salt et al., 1998; Mench et al., 2009)

Phytotechnologies



- TEs
- Microorganisms
- * Volatile compounds
- Less toxic compounds

Trace elements - Contamination - Phytoremediation



Trace elements - Contamination - Phytoremediation

MAJOR CHALLENGES

1. **Conditioning of the soils**
 - **Acidity/alkalinity, fertility, structure**
2. **Plant survival & growth: species selection**
 - **Tolerance, adapted, accumulation/exclusion**
3. **Monitoring and assessment**
 - **Eco-toxicity, bioindicators, ecosystem services**
4. **Management of the generated biomass**
 - **Valorization options**

Conditioning of the soils



Soil amelioration and conditioning

Waste materials: organic and inorganic

Recycling & reutilization

End of waste

Circular economy

Amendments selection: crucial

Allow/Improve plants growth

TEs solubility, mobility, speciation & bioavailability

Immobilization

↓ solubility/availability

Combination of amendments

Extraction

↑ solubility/availability

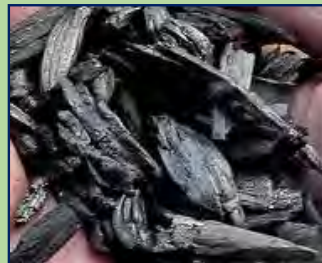
Acid, fresh/redox active



Conditioning of the soils

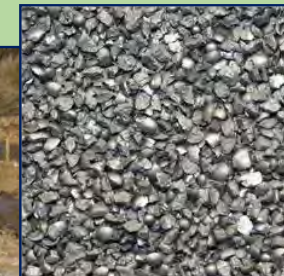
ORGANIC AMENDMENTS

- ✓ Sewage sludge
- ✓ Municipal waste
- ✓ Manure
- ✓ Compost
- ✓ Agri-food byproducts
- ✓ Biochar
- ✓ Digestate



INORGANIC AMENDMENTS

- ✓ Lime
- ✓ Zeolites
- ✓ Fe rich materials
- ✓ Nanoparticles
- ✓ Red mud
- ✓ Other residues (paper mill)



Trace elements - Contamination - Phytoremediation

MAJOR CHALLENGES

1. Conditioning of the soils

- Acidity/alkalinity, fertility, structure

2. Plant survival & growth: species selection

- Tolerance, adapted, accumulation/exclusion

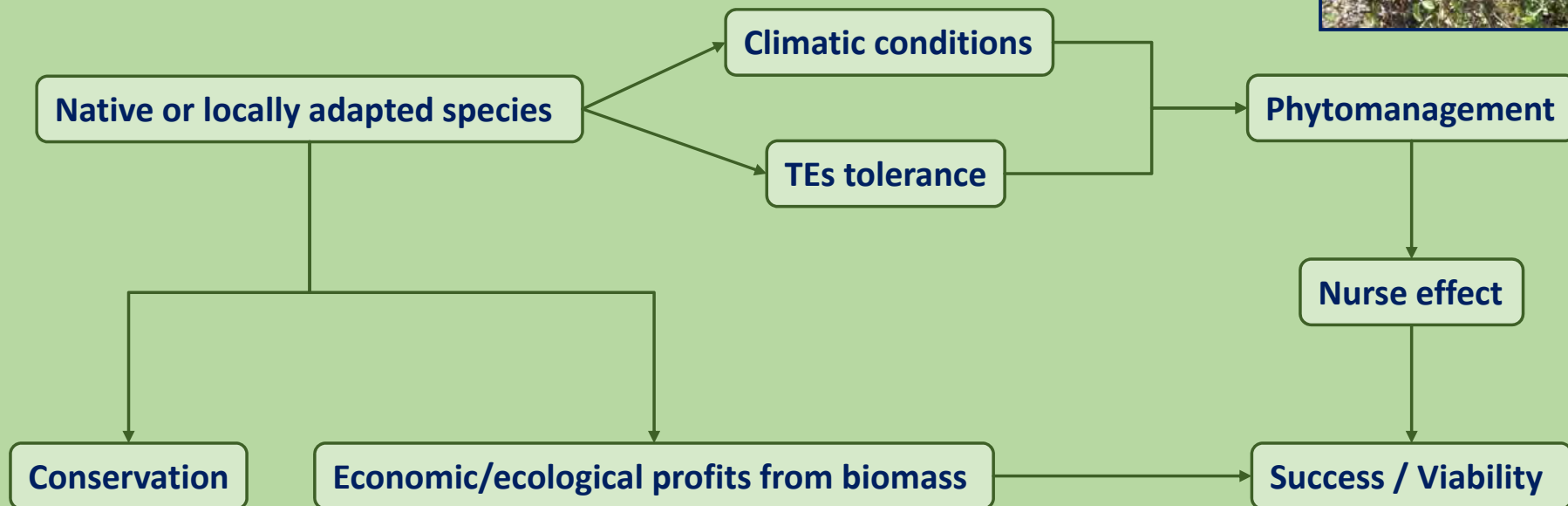
3. Monitoring and assessment

- Eco-toxicity, bioindicators, ecosystem services

4. Management of the generated biomass

- Valorization options

Selection of the plants



Selection of the plants: extraction vs stabilisation



Pteris vittata L. (As accumulator;
Cao et al., 2003)



Nocaea caerulescens (Cd & Zn
hyperaccumulator; Baker et al., 1994)



Alyssum murale (Ni & Cd
phytomining; Chaney et al., 1999)

Agrostis capilaris, *Festuca rubra*
(Vangronsveld et al., 1998)



Soya, rapeseed,
sunflower, sorghum,
common cane
(economic revenue)



Salix spp. (willow) y *Populus*
spp. (poplar) (high biomass,
fast growth, energy crops)



Selection of the plants: native/adapted species



Dittrichia viscosa (L.) Greuter



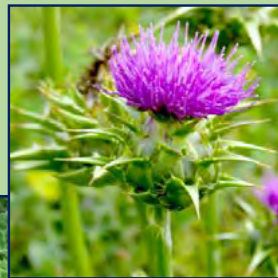
Atriplex halimus (L.)



Sylibum marianum (L.) Gaertn



Piptatherum miliaceum (L.)
Cosson



Bituminaria bituminosa (L.)
Stirton

Nicotiana glauca Graham



Trace elements - Contamination - Phytoremediation

MAJOR CHALLENGES

1. Conditioning of the soils

- Acidity/alkalinity, fertility, structure

2. Plant survival & growth: species selection

- Tolerance, adapted, accumulation/exclusion

3. Monitoring and assessment

- Eco-toxicity, bioindicators, ecosystem services

4. Management of the generated biomass

- Valorization options

Monitoring and assessment

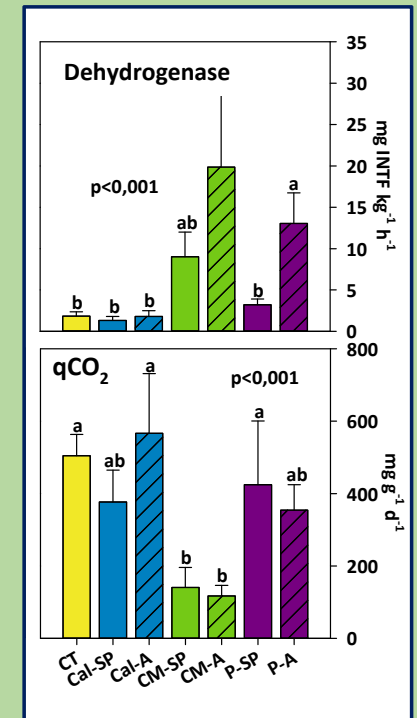
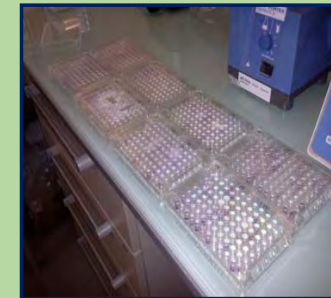
SOIL HEALTH

Indicators to monitor soil restoration:

- Trace elements solubility
- Soil solution (pore water)
- Accumulation in the plants

Bio-indicators (microbial parameters):

- ✓ Microbial community size (CO₂-C, Biomass-C, qCO₂)
- ✓ Biological functionality, general activity and functional diversity (enzymatic activities, FDA, CLPPs, etc.)

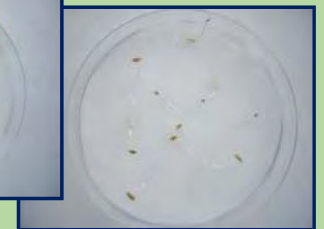
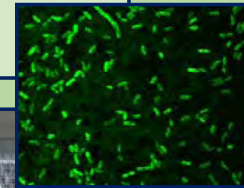


Monitoring and assessment

SOIL HEALTH

Eco-toxicity Tests (bio-assays):

- ✓ Direct (earthworms (*Eisenia fetida*) mortality, plant growth test)
- ✓ Indirect (*Vibrio fischeri* luminescence inhibition, *Daphnia magna*/*Thamnocephalus platyurus* immobilization, germination indices)



Trace elements - Contamination - Phytoremediation

MAJOR CHALLENGES

1. Conditioning of the soils

- Acidity/alkalinity, fertility, structure

2. Plant survival & growth: species selection

- Tolerance, adapted, accumulation/exclusion

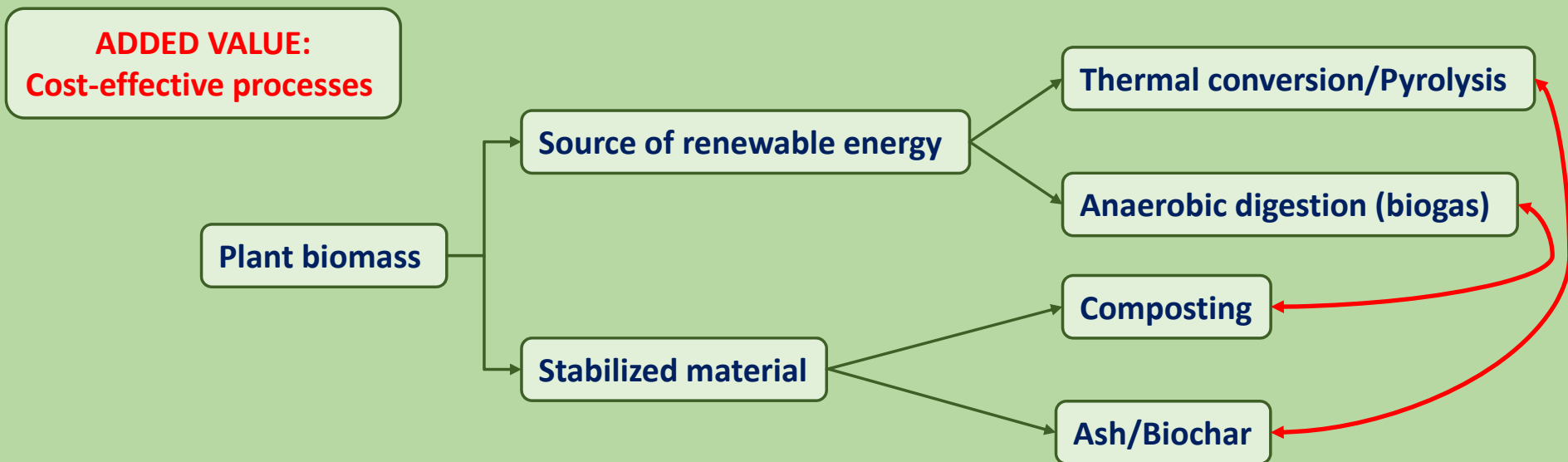
3. Monitoring and assessment

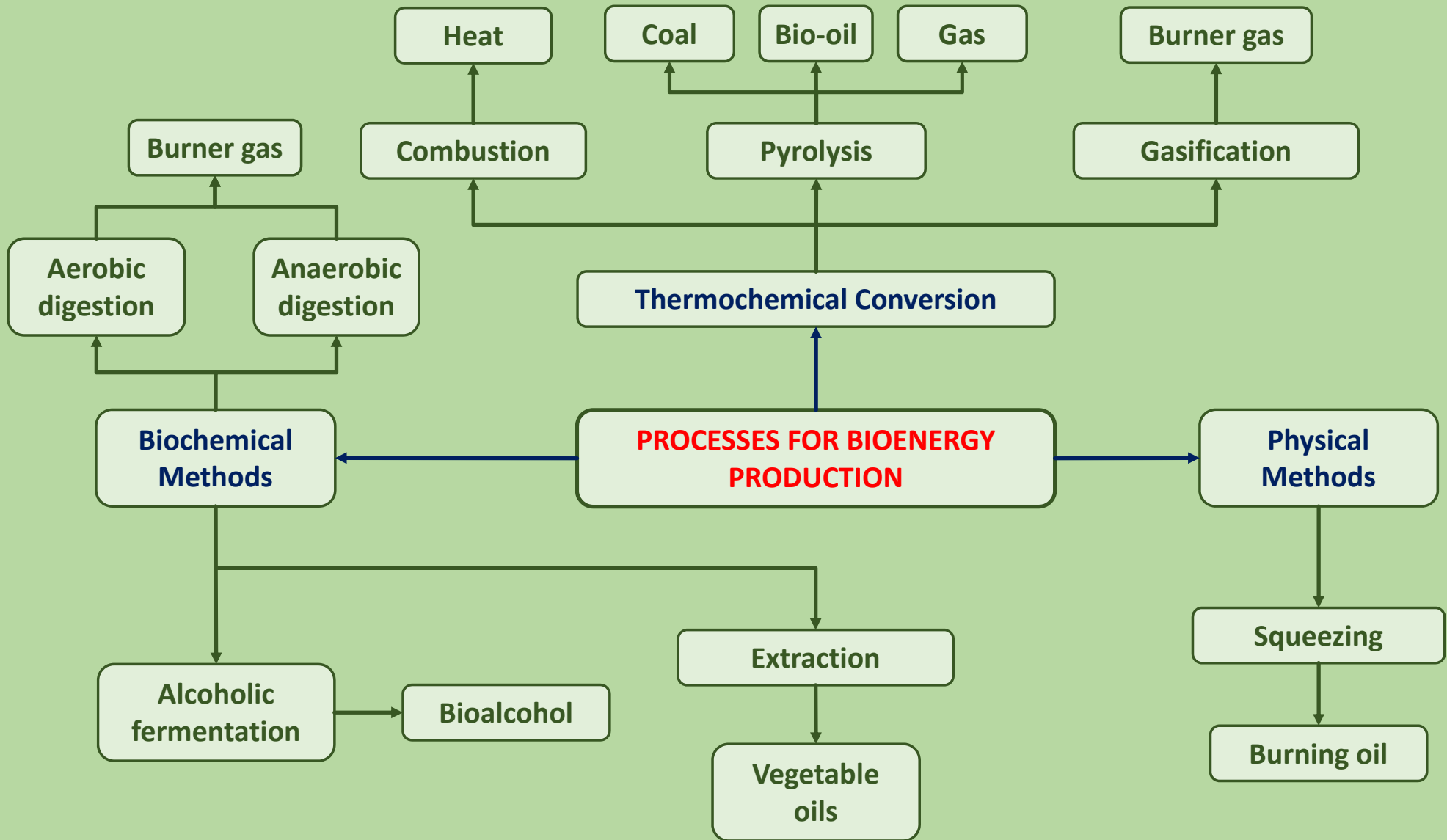
- Eco-toxicity, bioindicators, ecosystem services

4. Management of the generated biomass

- Valorization options

Management of the generated biomass





Management of the generated biomass

Biomass from phytoremediation

- ✓ Potential use as energy crops
- ✓ Degradability and biogas generation potential (ignocellulosic components/carbohydrate rich, high content of biodegradable organic matter)
- ✓ Biomass thermic potential (low water and ashes content; high amount of raw lignin)
- ✓ TEs effects on biomass energetic potential

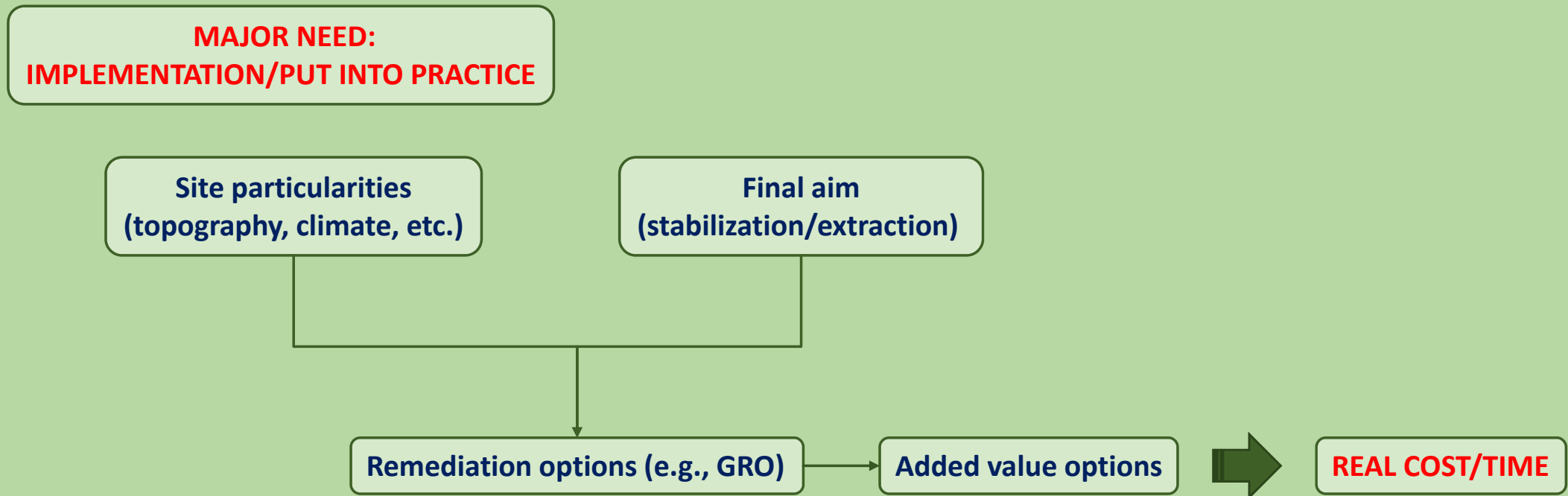
Other advantages

Impact on agricultural soil

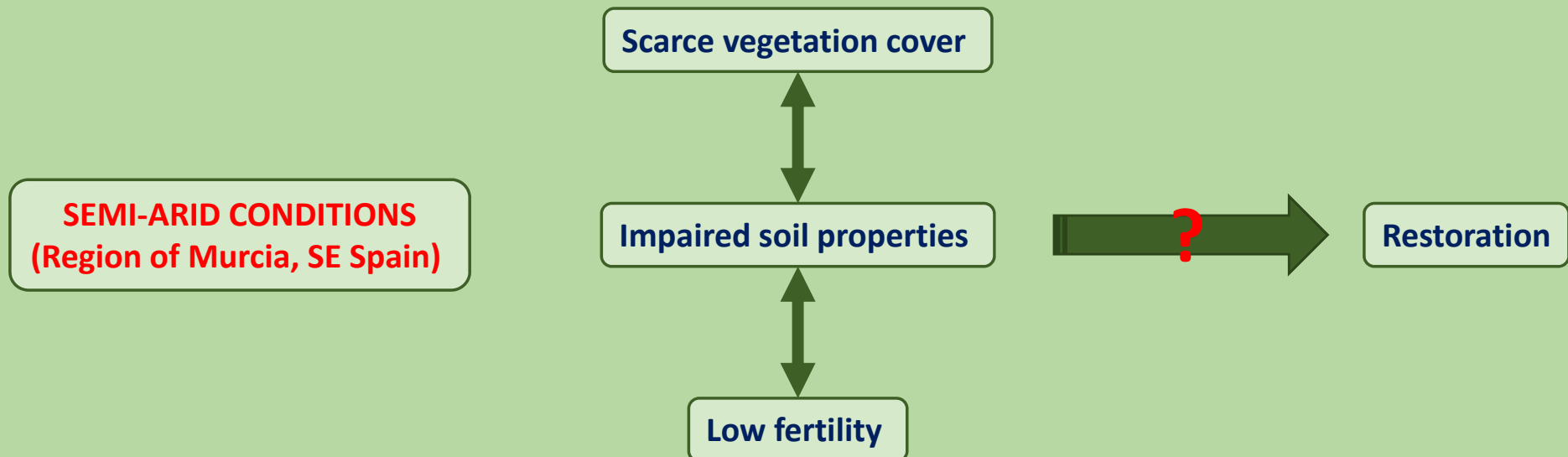
- ✓ (Indirect) Land use change
- ✓ Bio-fuel life cycle emissions ↓



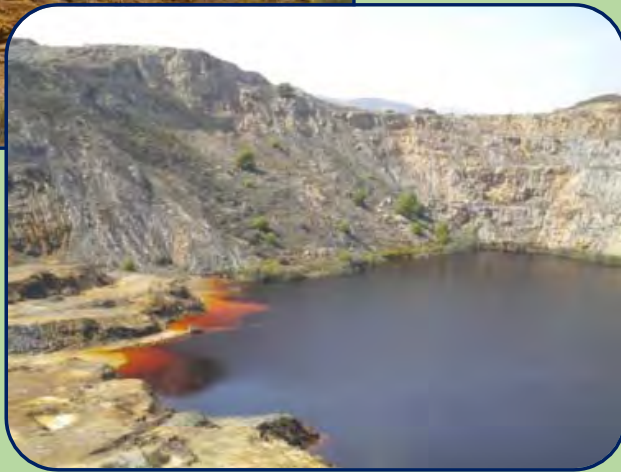
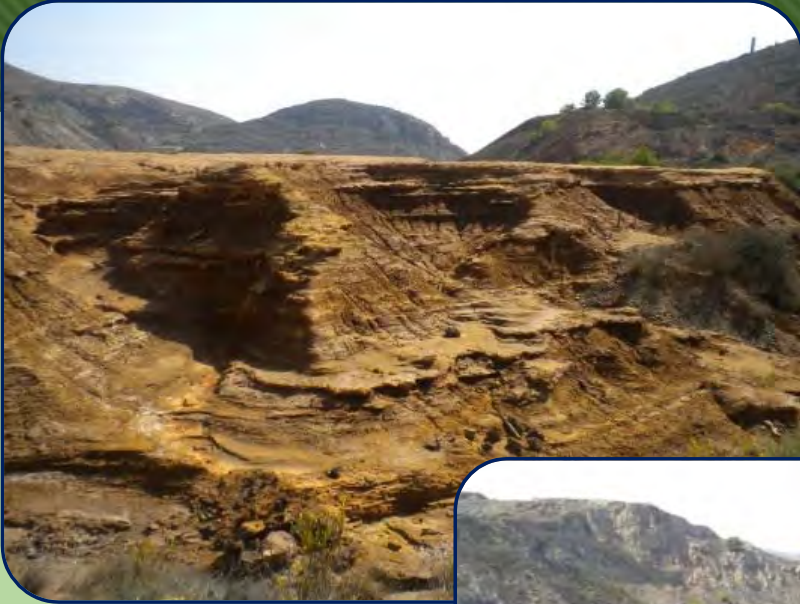
Trace elements - Contamination - Phytoremediation



Trace elements - Contamination - Phytoremediation



La Unión-Cartagena Mining Area (Murcia, Spain)



La Unión-Cartagena Mining Area (Murcia, Spain)



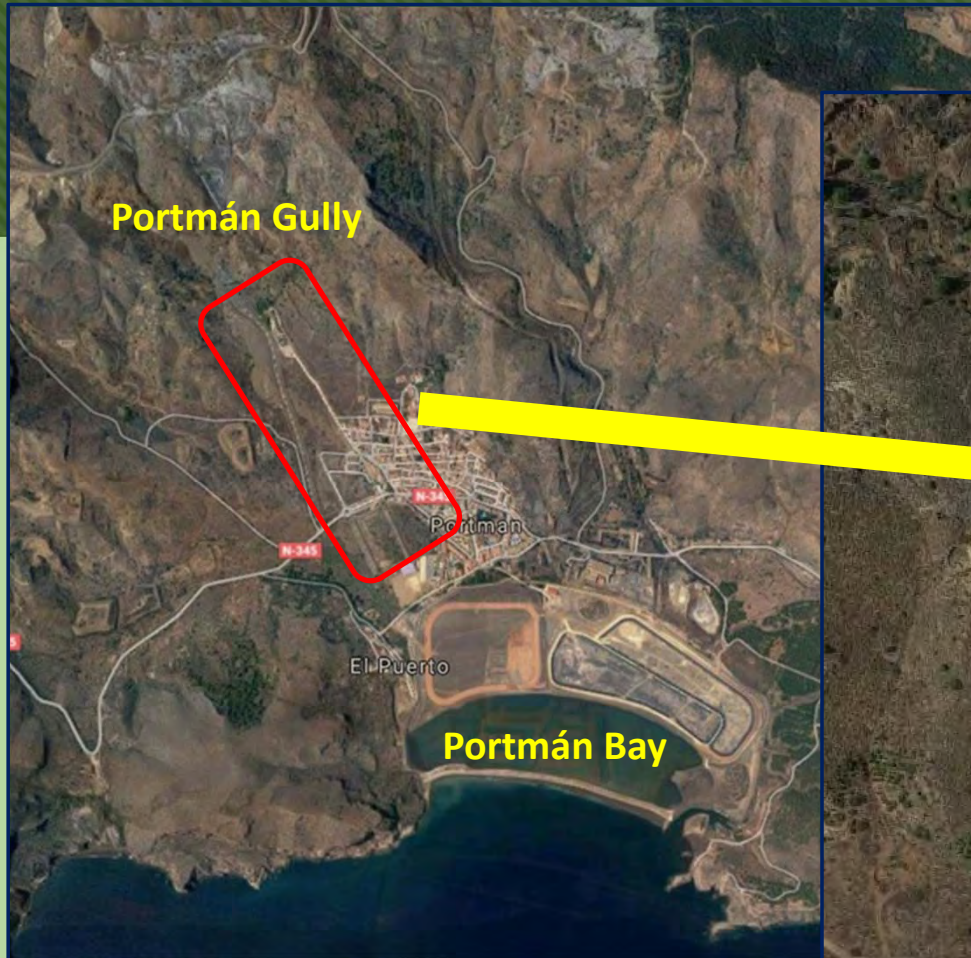
Case study 1 - **Options for mine site restoration**

- 1. Soil survey within the mining area of La Unión-Cartagena (Murcia, Spain)**
- 2. Selection of amendments**
- 3. Plant species: adapted, added value**
- 4. Pot test: performance of plants and amendments**
- 5. Upscale: biomass production, eco-toxicity**

Experimental site location



Experimental site location



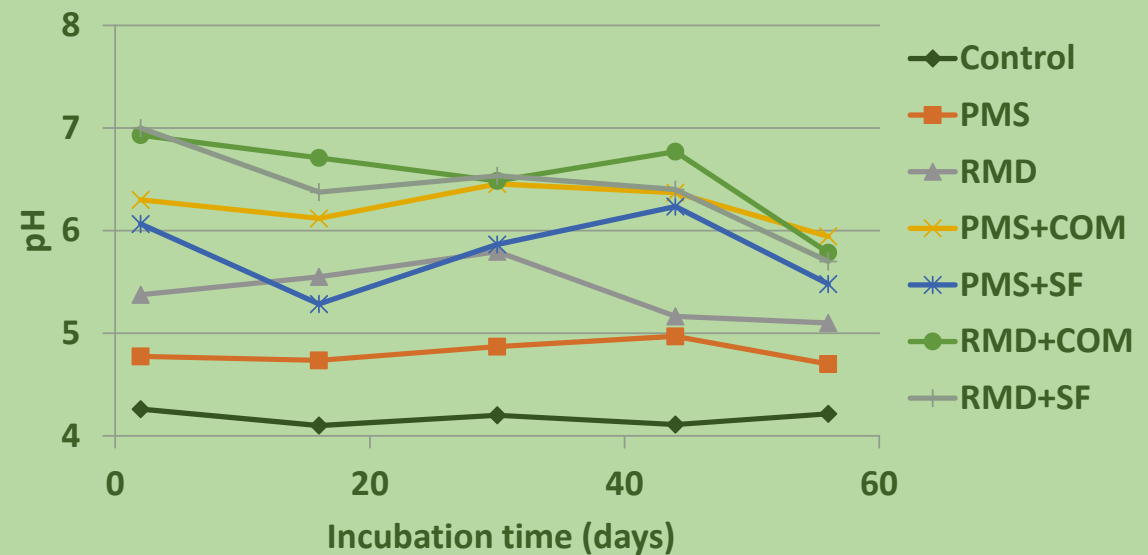


Soil characterisation and conditioning

	Portmán soil	CaCl ₂ /NaHCO ₃
pH	4.16	-
TOC (g kg ⁻¹)	2.7	-
EC (mS cm ⁻¹)	2.22	-
As (mg kg ⁻¹)	1976	0.67
Cd (mg kg ⁻¹)	12	0.2
Cu (mg kg ⁻¹)	230	1.6
Fe (mg kg ⁻¹)	214747	0.5
Mn (mg kg ⁻¹)	968	9.3
Pb (mg kg ⁻¹)	19129	1667
Zn (mg kg ⁻¹)	2257	28

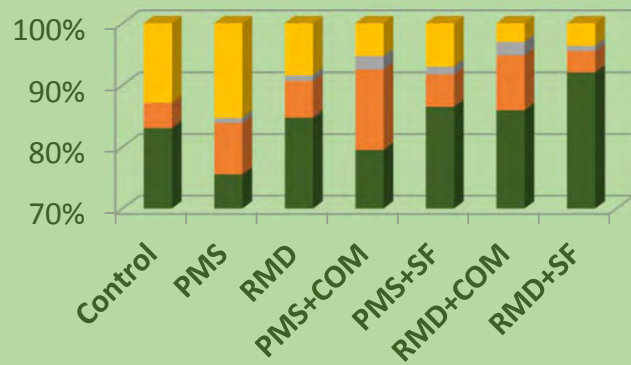


Paper mill sludge (PMS) - Red mud derivative (RMD)
Pig slurry (solid fraction, SF) - Compost (olive mil waste, COM)

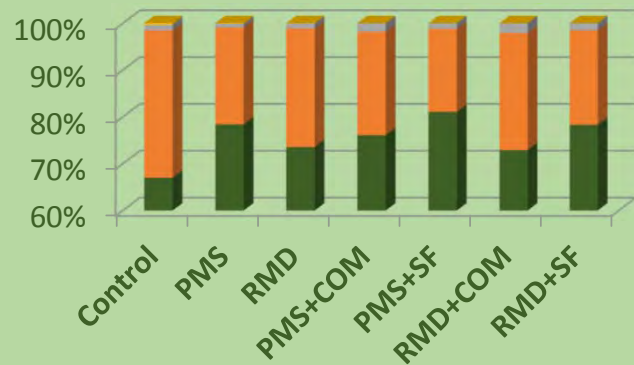


Sequential extractions

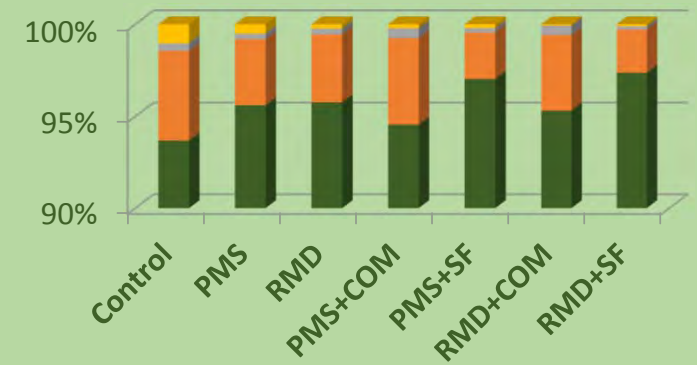
Cd



Pb



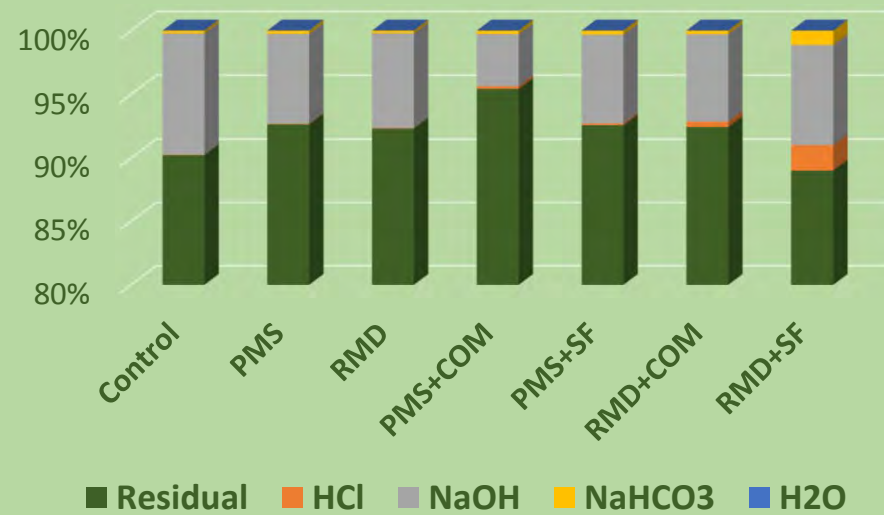
Zn



■ Residual ■ EDTA ■ NaOH ■ CaCl2

Sequential extractions

As



Plants test



Plants test



- ✓ *Cynara cardunculus* ↓
- ✓ *Silybum marianum* ↑
- ✓ *Nicotiana glauca* ↑
- ✓ *Limonium caesium* ↓
- ✓ *Piptatherum miliaceum* ↑

Pot test

Soil and amendments

	Soil	SF	PMS	RMD
pH	3.80	7.31	8.87	10.9
EC (dS m ⁻¹)	0.19	6.85	4.40	0.14
OM (%)	0.28	68.6	32.2	n.d.
TOC (g kg ⁻¹)	1.63	356	195	7.17
TN (g kg ⁻¹)	0.31	32.8	20.0	0.15
P (g kg ⁻¹)	0.29	7.7	0.26	0.37
As (mg kg ⁻¹)	1976	<0.1	<0.1	4.68
Cd (mg kg ⁻¹)	12.0	0.2	0.19	0.27
Cu (mg kg ⁻¹)	230	226	184	55.7
Fe (g kg ⁻¹)	215	1.36	9.10	90.2
Mn (mg kg ⁻¹)	968	194	72.7	901
Pb (mg kg ⁻¹)	19129	5.3	18.4	41.2
Zn (mg kg ⁻¹)	2257	849	38.6	10.7

n.d.: not determined.

- ✓ Pig slurry (solid fraction, SF)
- ✓ Paper mill sludge (PMS)
- ✓ Red mud derivative (RMD)

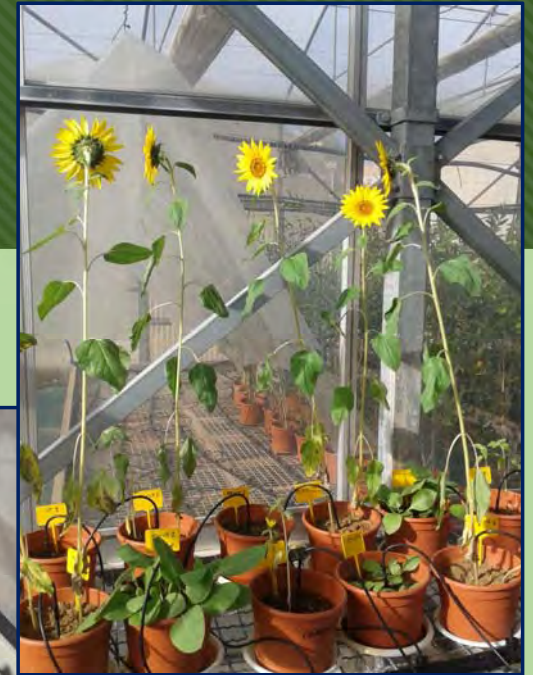
Pot test

S. marianum
P. miliaceum



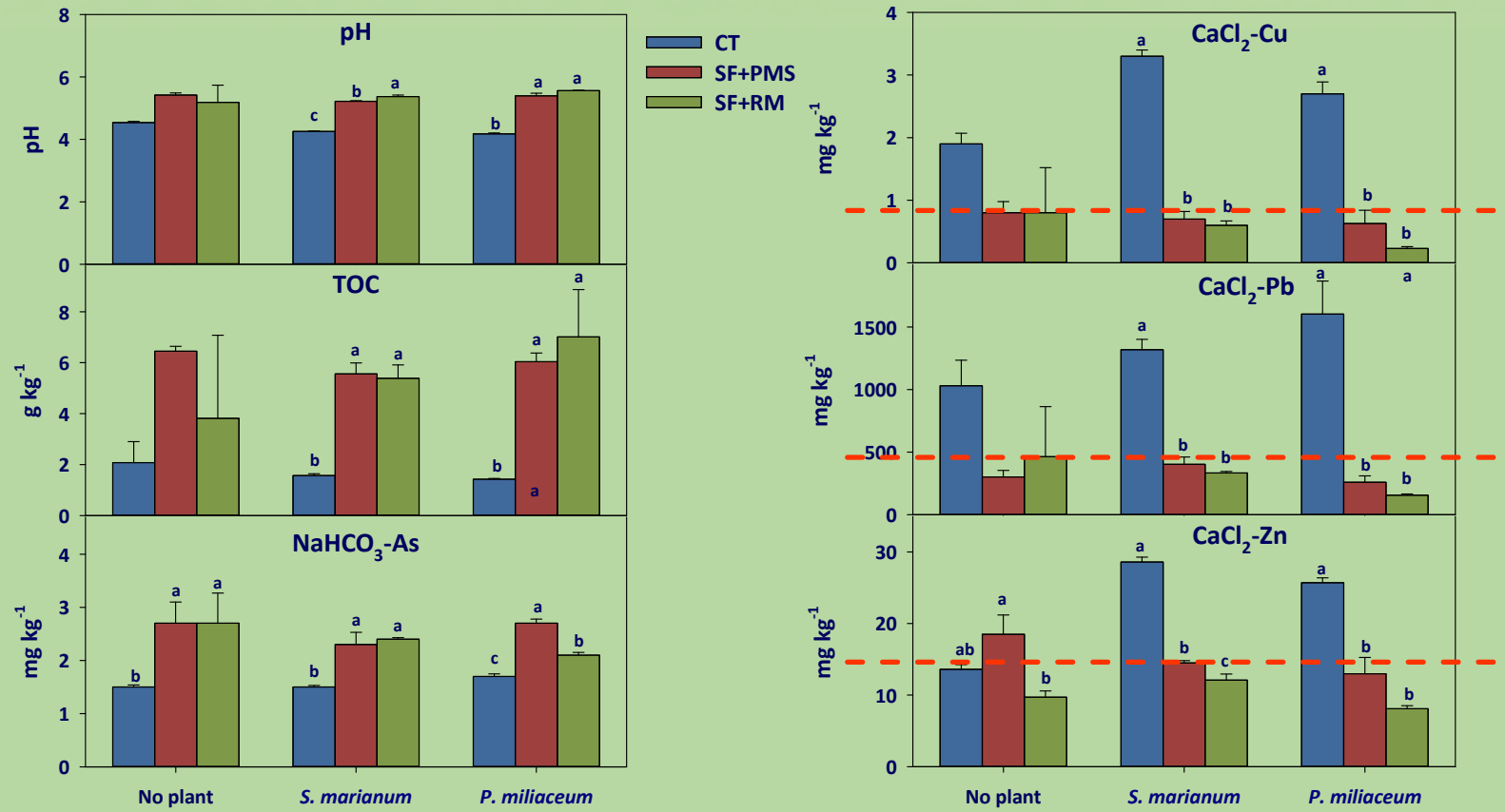
Pot test

N. glauca
H. annuus



Pot test Effects on soils

pH, TOC,
Metals & As extractability



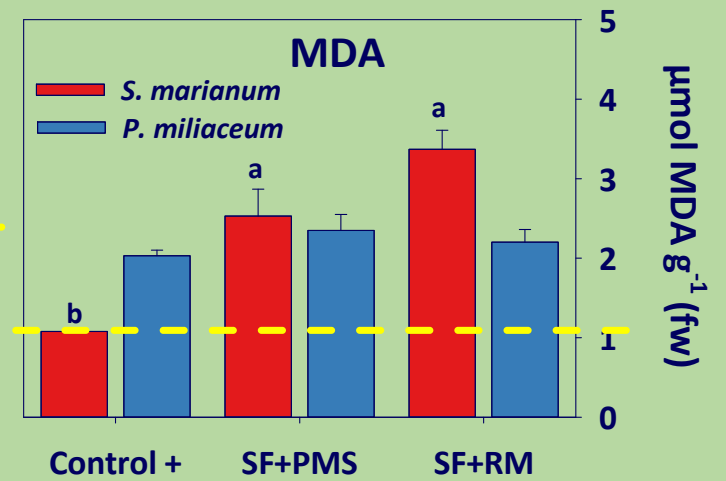
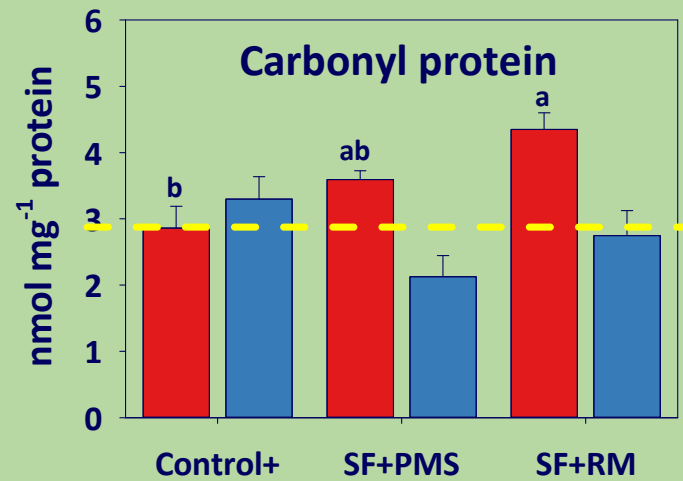
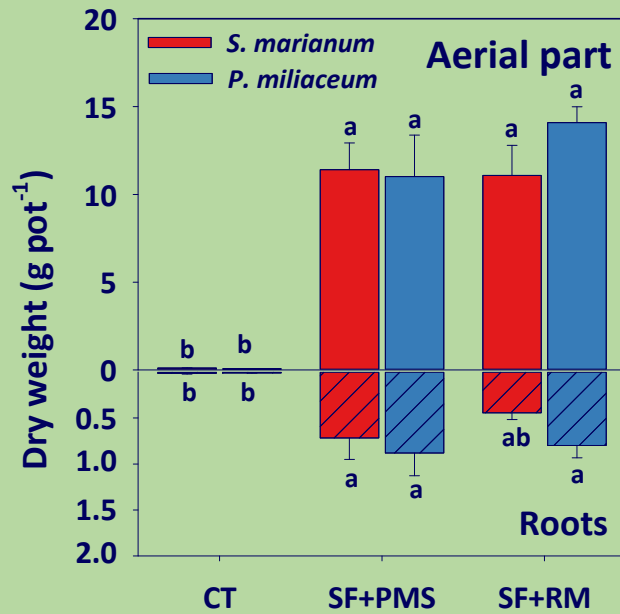
Pot test Plants – TEs accumulation

TE CONCENTRATIONS (mg kg⁻¹ dw) IN THE PLANTS (mean ± se)

	As	Cd	Cr	Cu	Pb	Zn	Mn
<i>Piptatherum miliaceum</i>							
RM	7.3 ± 1.20	0.05 ± 0.10	63.6 ± 9.5ab	20.0 ± 1.0b	91 ± 3.0	124 ± 2.5cd	211 ± 8.0d
PMS	8.4 ± 2.45	0.03 ± 0.02	25.1 ± 4.4b	19.0 ± 1.2b	114 ± 24.5	99 ± 0.1d	168 ± 10.0cd
<i>Silybum marianum</i>							
RM	2.5 ± 0.60	1.95 ± 0.15	73.5 ± 8.5ab	21.3 ± 0.8b	118 ± 18.5	293 ± 17.0b	639 ± 4.0a
PMS	3.2 ± 0.90	2.50 ± 0.50	123.0 ± 40a	27.8 ± 0.6a	183 ± 10.5	407 ± 12.5a	577 ± 6.5b
<i>Helianthus annuus</i>							
RM	0.6 ± 0.52	0.45 ± 0.05	12.4 ± 0.91b	6.3 ± 0.4d	87 ± 10.0	100 ± 7.0d	229 ± 22.5c
PMS	1.2 ± 0.15	0.59 ± 0.05	24.7 ± 5.20b	7.9 ± 1.5d	161 ± 5.5	145 ± 6.9c	239 ± 8.0c
<i>Nicotiana glauca</i>							
RM	2.6 ± 1.76	1.50 ± 0.06	21.8 ± 2.48b	10.0 ± 1.1c	225 ± 6.5	137 ± 9.5c	72.5 ± 12.0e
PMS	1.6 ± 0.37	1.59 ± 0.36	18.1 ± 1.87b	10.1 ± 0.9c	231 ± 26	137 ± 3.0c	78.0 ± 5.0e
ANOVA							
TxP	n.s.	n.s.	*	**	n.s.	**	**

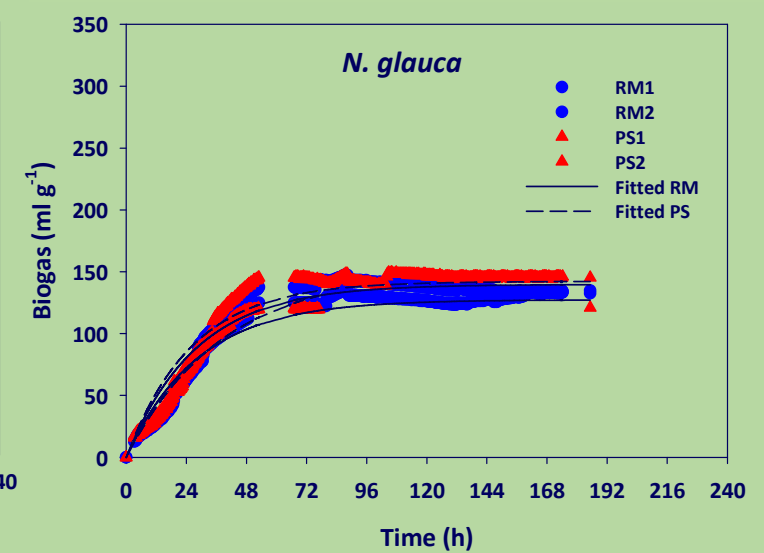
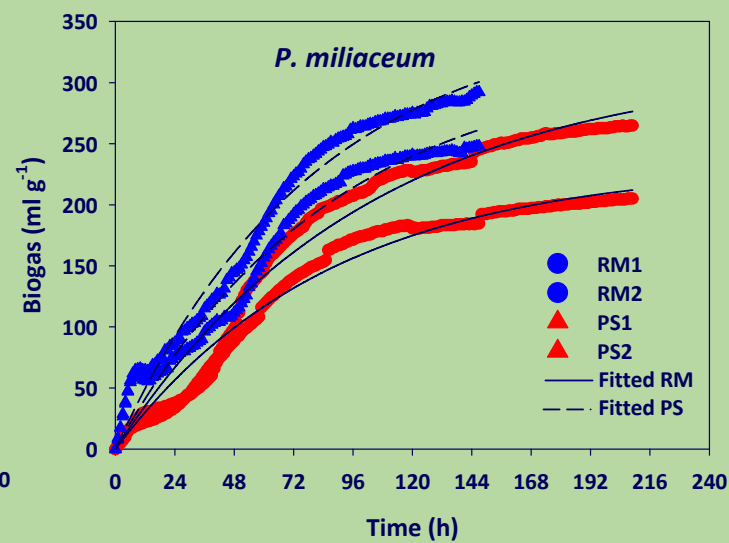
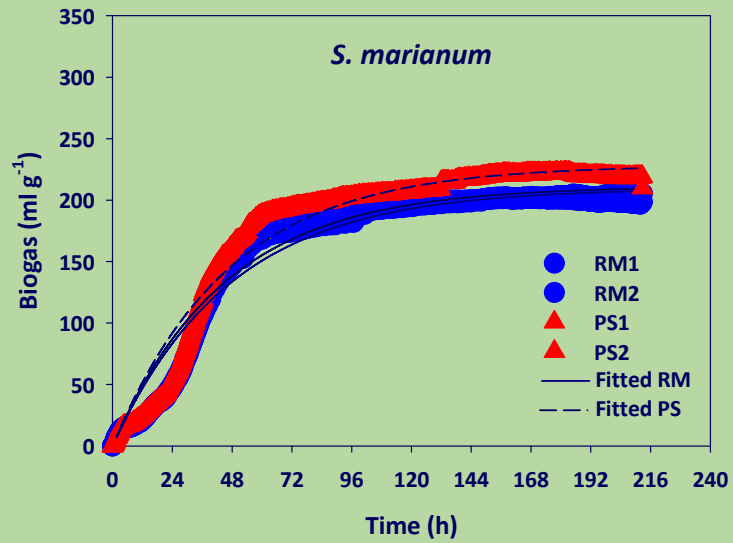
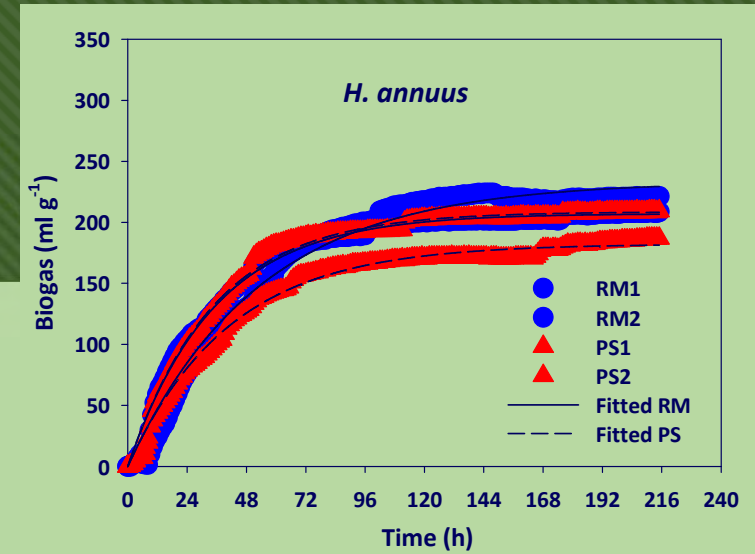
*, **: $P < 0.05$, and 0.01 , respectively; n.s.: not significant. Post-hoc: Tukey's test at $P < 0.05$.

Pot test Plants - Oxidative stress



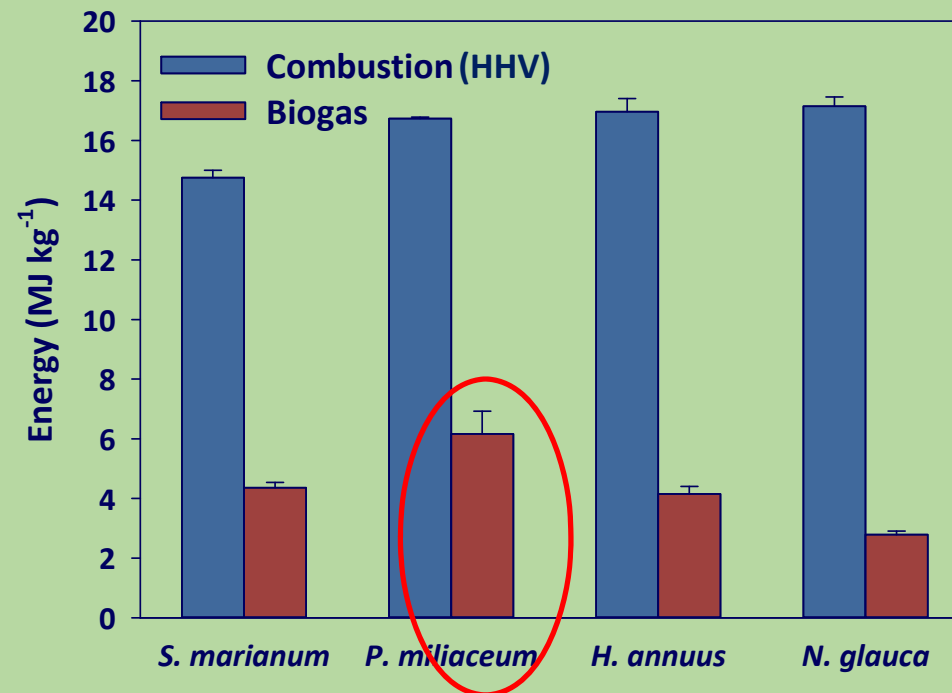
Pot test Biomass valorisation

ANAEROBIC INCUBATION (ANKOM RF) - BIOGAS



Pot test Biomass valorisation

HHV: Higher heating value (calculated from elemental composition)



Planter test

Silybum marianum
Helianthus annuus

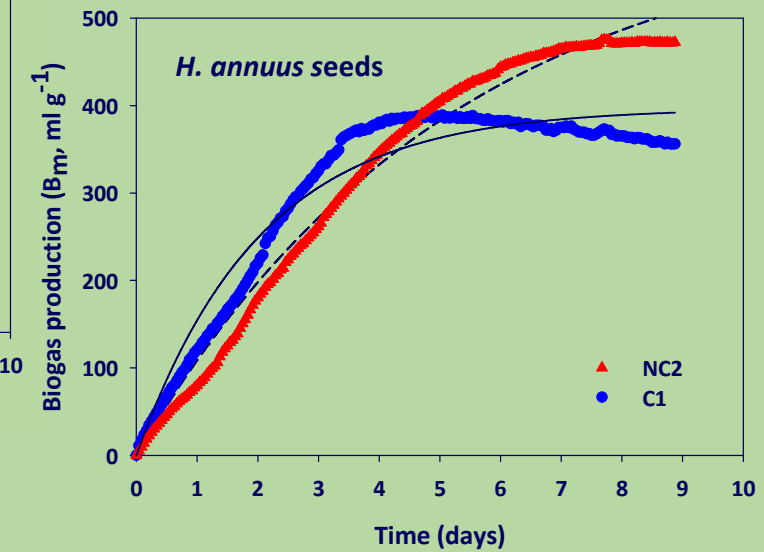
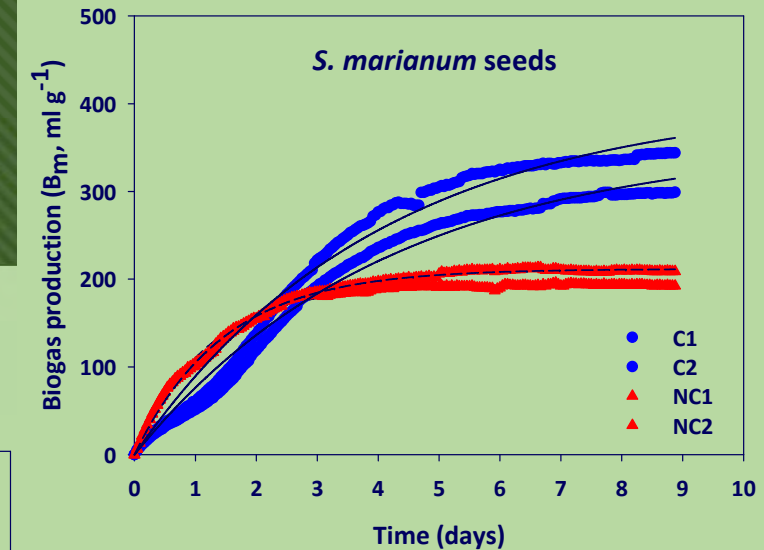
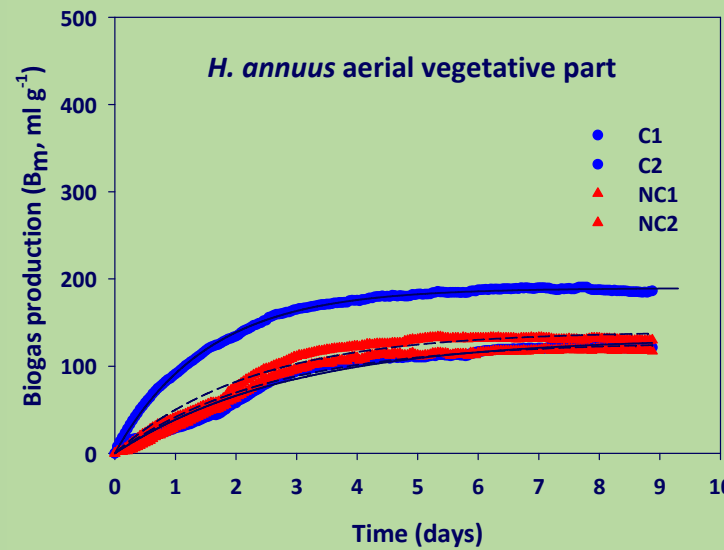
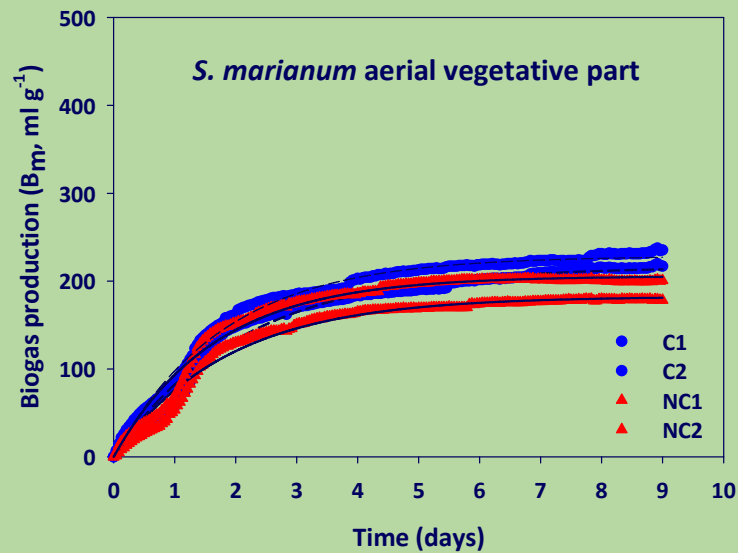
CONTAMINATED SOIL
SF + PMS

NON CONTAMINATED SOIL



Planter test **Biomass valorisation**

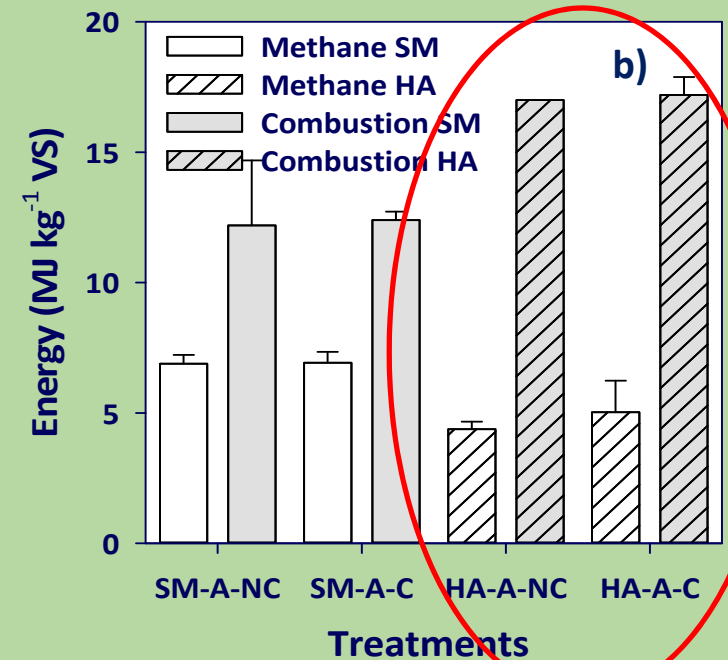
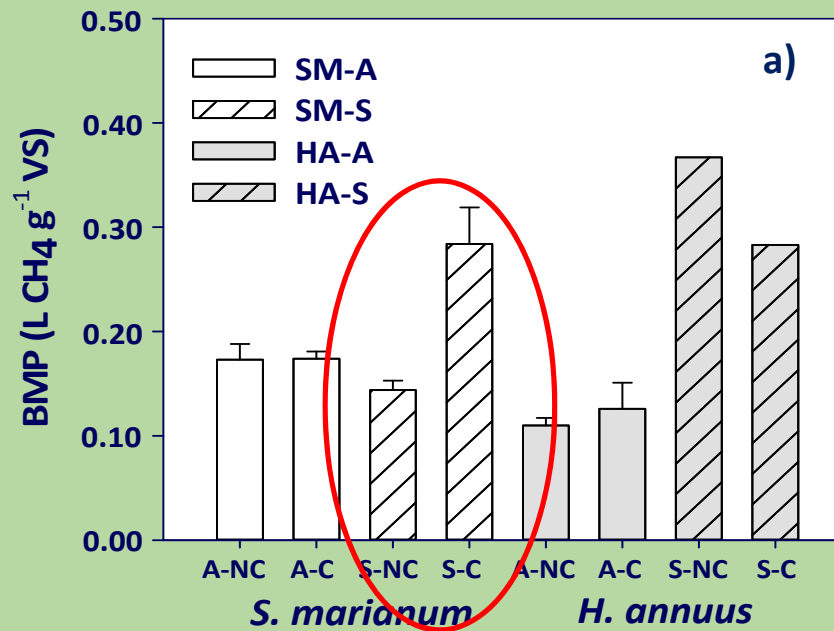
ANAEROBIC INCUBATION - BIOGAS



Planter test Biomass valorisation

a) Biomethane production potentials (BMP)

b) Comparison of energy production methods: anaerobic digestion (methane) and combustion (HHV)



Hunce et al., 2019. *Industrial Crops & Products* 135, 206-216.

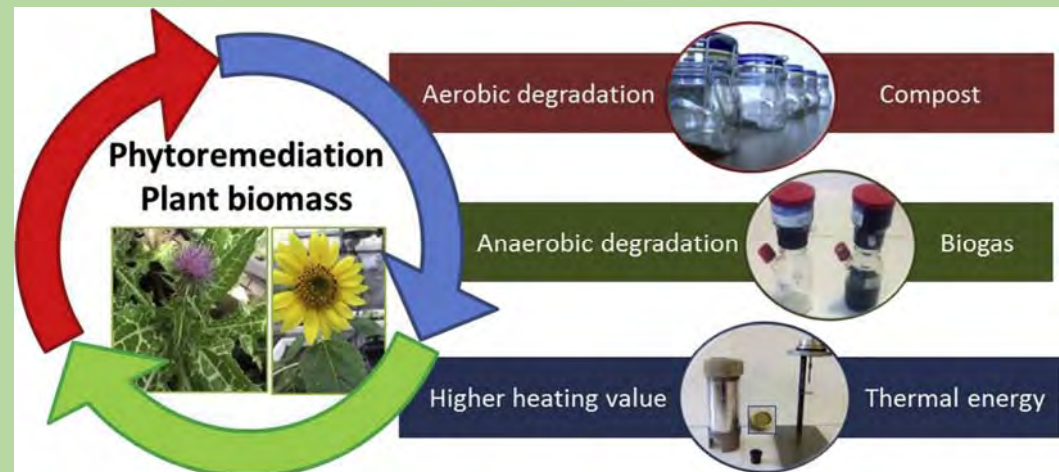
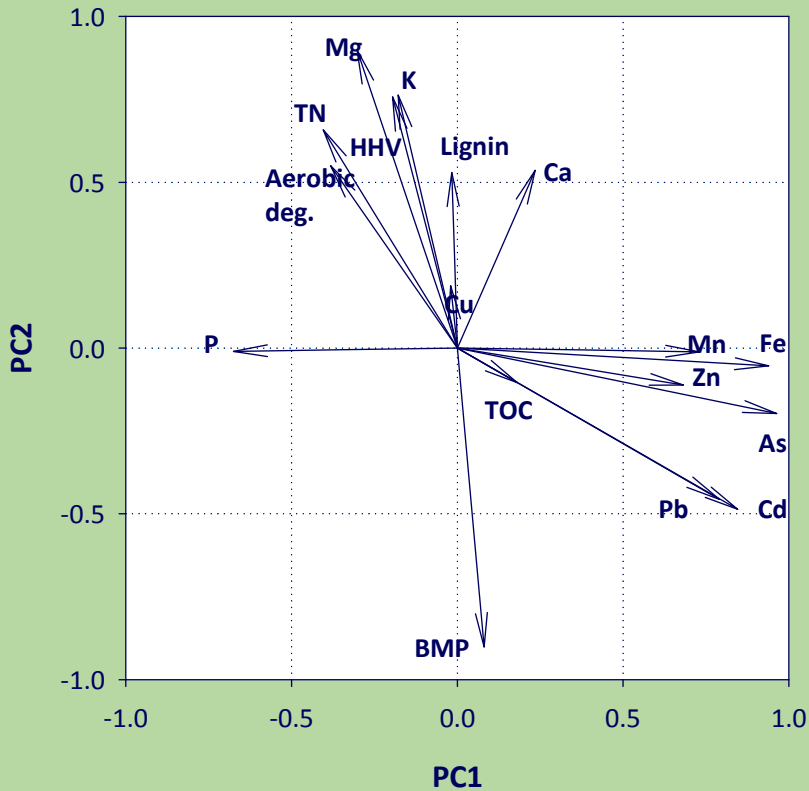
SM: *S. marianum*;
HA: *H. annuus*;

A: aerial part;
S: seeds;

NC: non-contaminated
C: contaminated

Planter test **Principal Component Analysis**

- ✓ **BMP (inversely), HHV and aerobic degradation related to lignin and nutrient content in the plants**
- ✓ **Metals and As concentration do not affect degradability and energy production potential**



Hunce et al., 2019. *Industrial Crops & Products* 135, 206-216.

Planter test **Eco-toxicological evaluation**

Plant growth test (direct) - *Helianthus annuus* & *Lactuca sativa*

Artificial soil (OCDE)

0.6 kg peat
1.6 kg clay
5.6 kg sand
0.08 kg CaCO₃



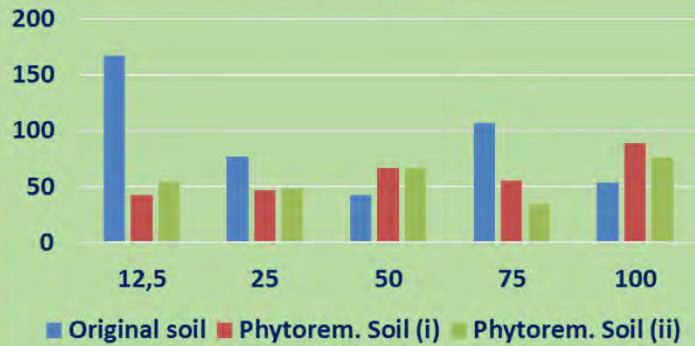
Soil dilutions



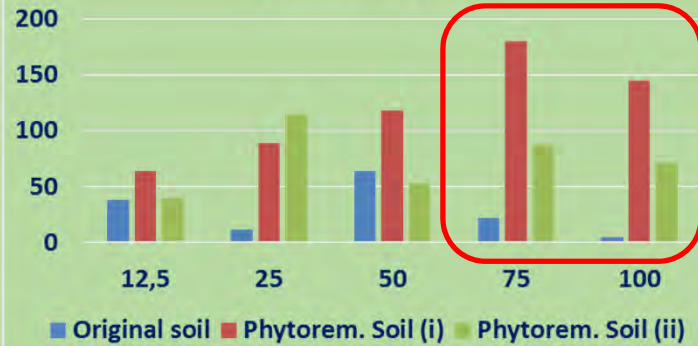
Helianthus annuus/*Lactuca sativa*
cultivated in growth chamber

Planter test **Eco-toxicological evaluation**

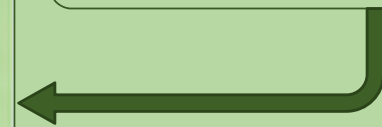
Sunflower (%FW)



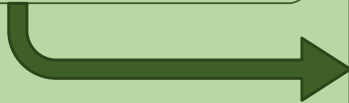
Lettuce (%FW)



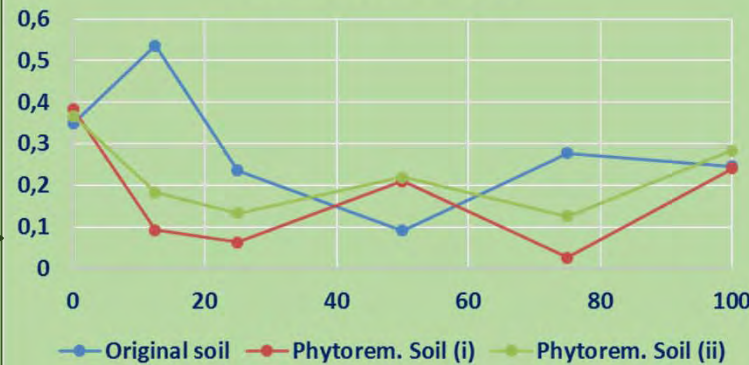
%Fresh weight compared to artificial (clean) soil



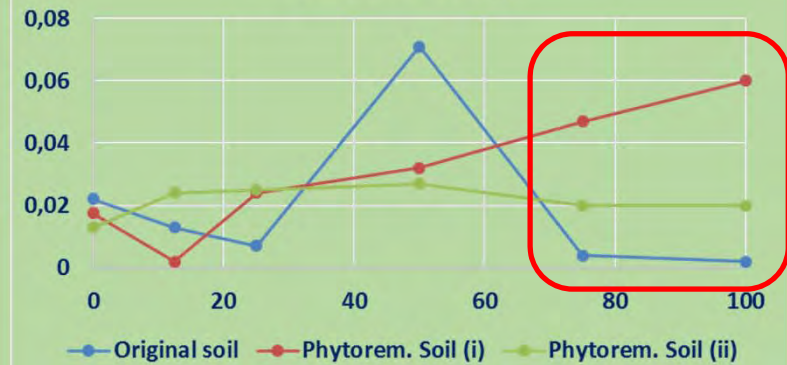
Total dry weight per soil dilution



Sunflower DW (g)

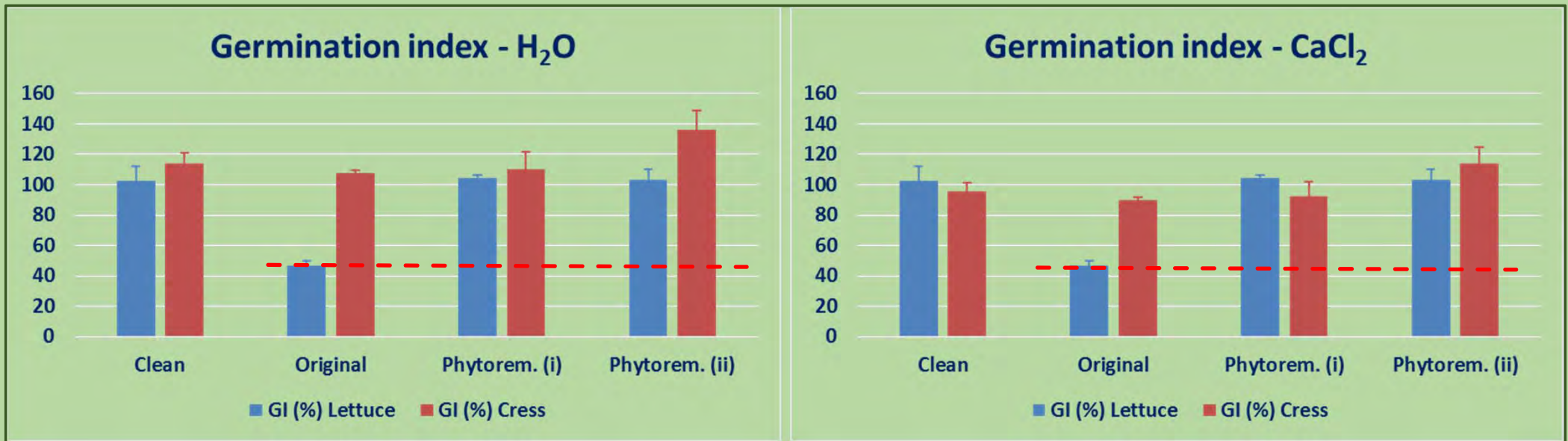
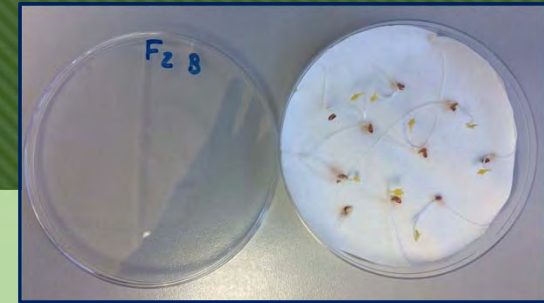


Lettuce DW (g)



Planter test **Eco-toxicological evaluation**

Germination index (indirect) - *Lepidium sativum* & *Lactuca sativa*

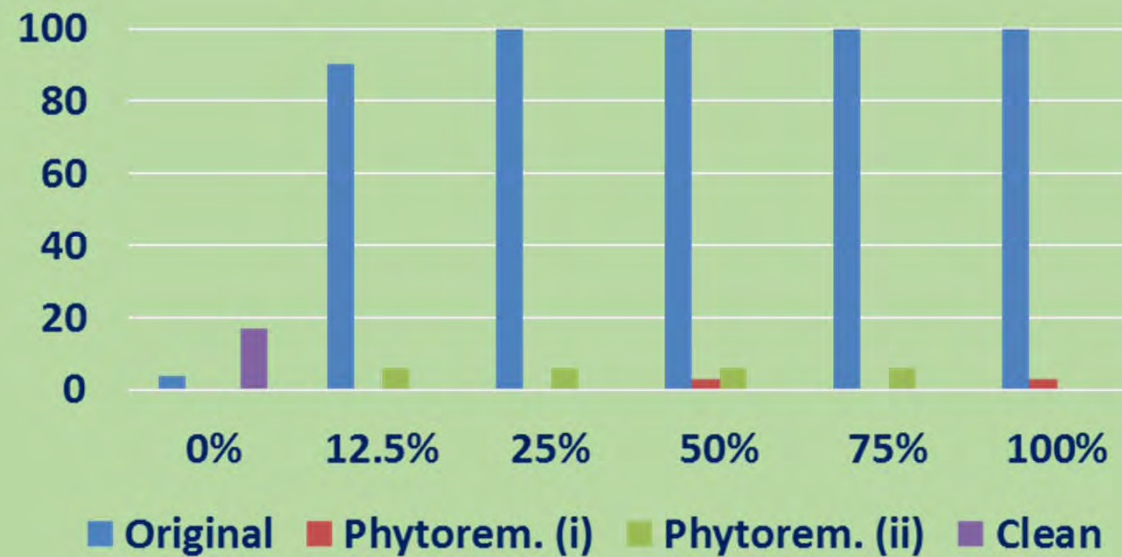


Planter test **Eco-toxicological evaluation**

Mortality test (indirect) – *Thamnocephalus platyurus*



% Mortality - *T. platyurus*



Case study 1 - CONCLUSIONS

- ✓ SF + inorganic amendments **stabilized TEs** in a mine soil
- ✓ Plants TE concentrations within common ranges for mine soils
- ✓ Carbonyl proteins and MDA levels confirmed oxidative stress in plants
- ✓ Amendments-native species combination suitable for mine soils
phytostabilization
- ✓ **No effects of TEs** in the plants on bioenergy production were observed
- ✓ Combustion was the best option for **bioenergy production** from these plants
- ✓ Phytostabilized soils showed **no toxicity**: successful remediation

Case study 2 - Phytostabilization of a mine affected soil



Case study 2 - **Phytostabilization of a mine affected soil**



Case study 2 - Phytostabilization of a mine affected soil



	Soil LL
pH	6.18 ± 0.05
OM (%)	0.23 ± 0.02
TOC (g kg⁻¹)	1.35 ± 0.10
N_T (g kg⁻¹)	0.45 ± 0.07
Fe (g kg⁻¹)	108 ± 1
Zn (mg kg⁻¹)	9686 ± 252
Pb (mg kg⁻¹)	10,188 ± 97
Cu (mg kg⁻¹)	193 ± 8
Mn (mg kg⁻¹)	4073 ± 368
Cd (mg kg⁻¹)	19 ± 1
As (mg kg⁻¹)	664 ± 28

Case study 2 - Phytostabilization of a mine affected soil



PIG SLURRY
(60 m³ ha⁻¹)



HYDRATED LIME
(>93% CaO)
(2.3 t ha⁻¹)



COMPOST
(60 t ha⁻¹)



Case study 2 - **Phytostabilization of a mine affected soil**

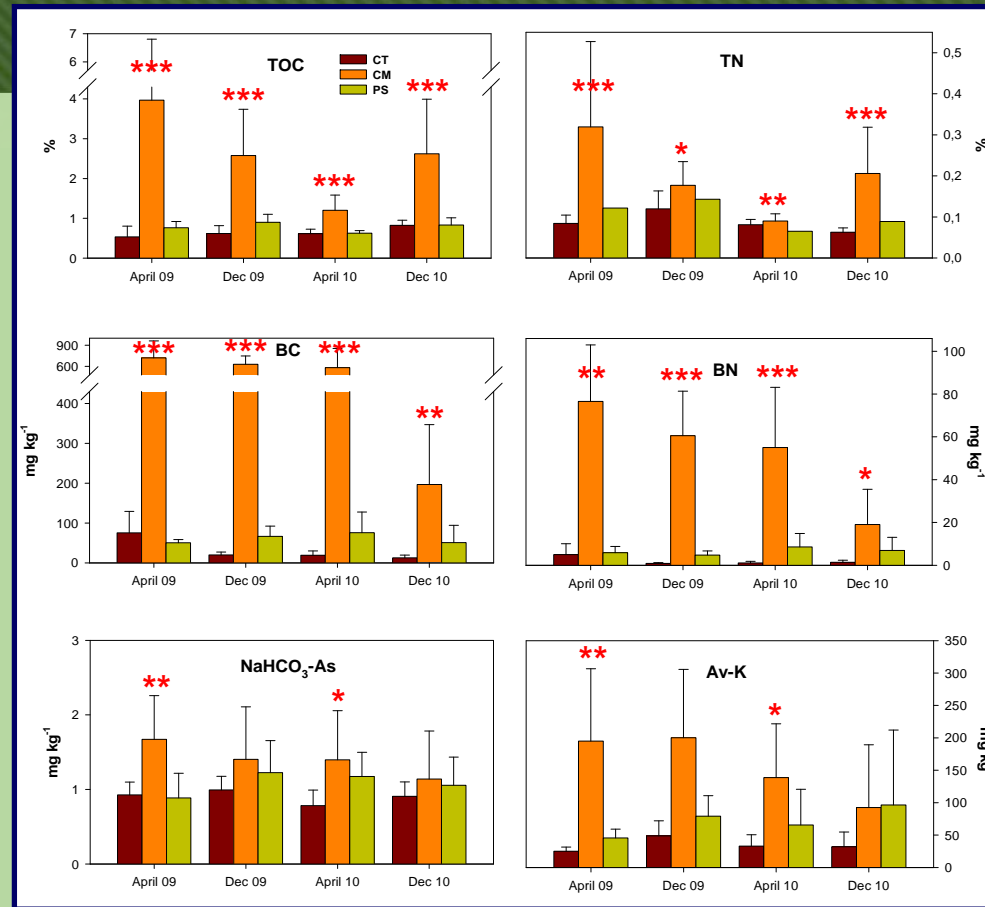


Atriplex halimus L.

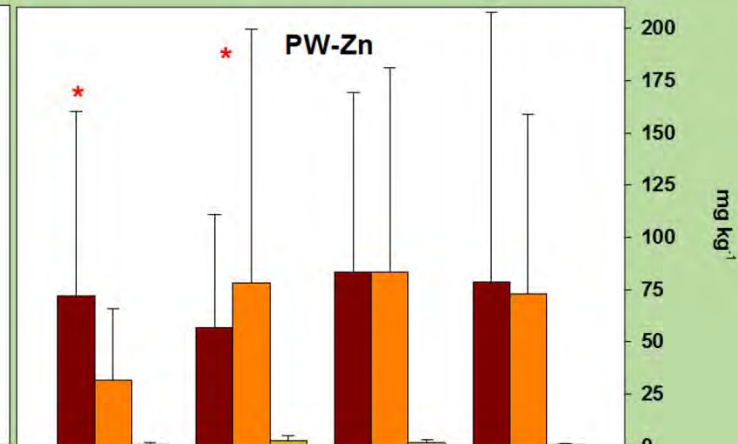
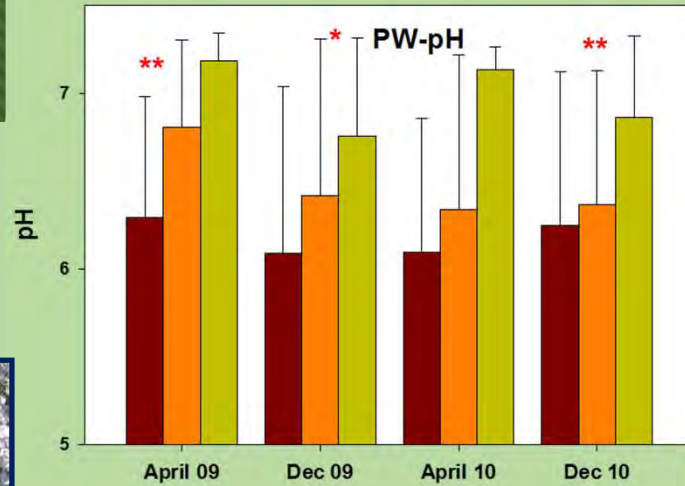


~~*Bituminaria bituminosa* L.~~

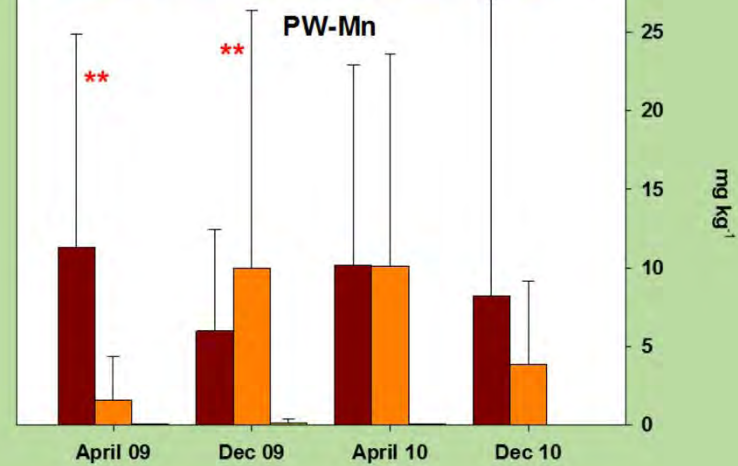
Case study 2 - SOILS



Case study 2 – PORE WATER

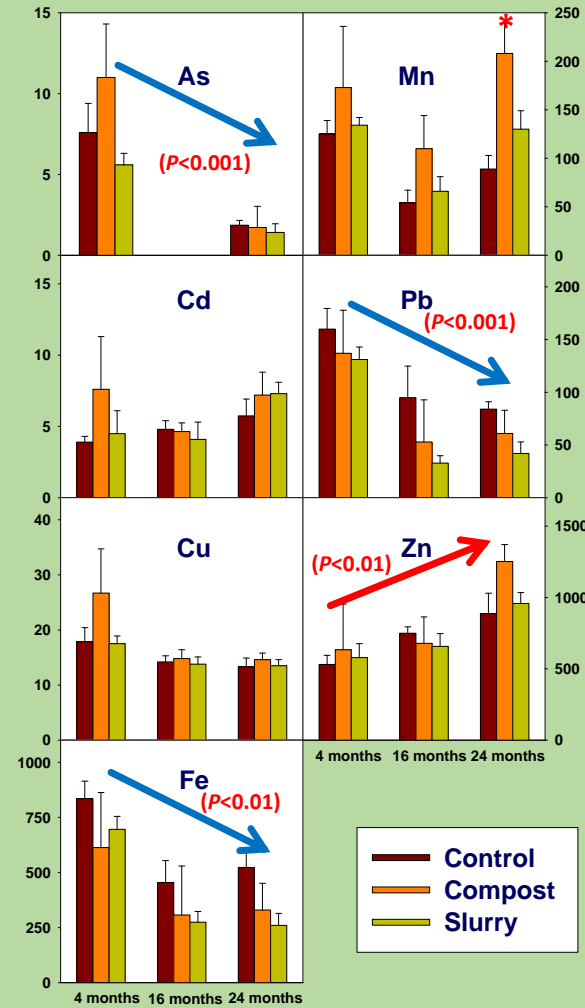


■ Control
■ Compost
■ Slurry



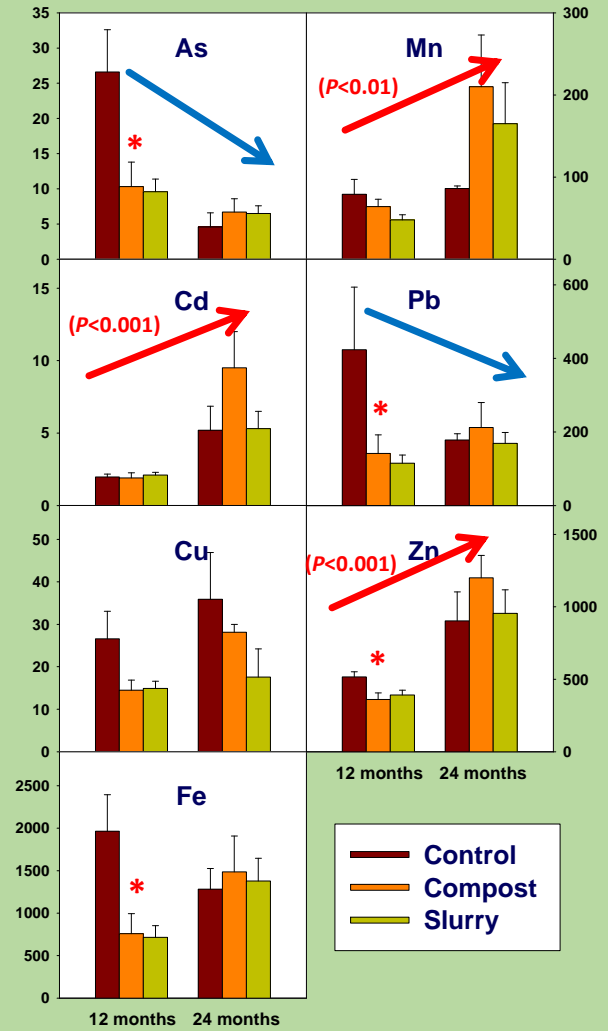
Case study 2

PLANTS



LEAF (mg kg⁻¹)

FRUIT (mg kg⁻¹)



Case study 2 - **PLANTS**

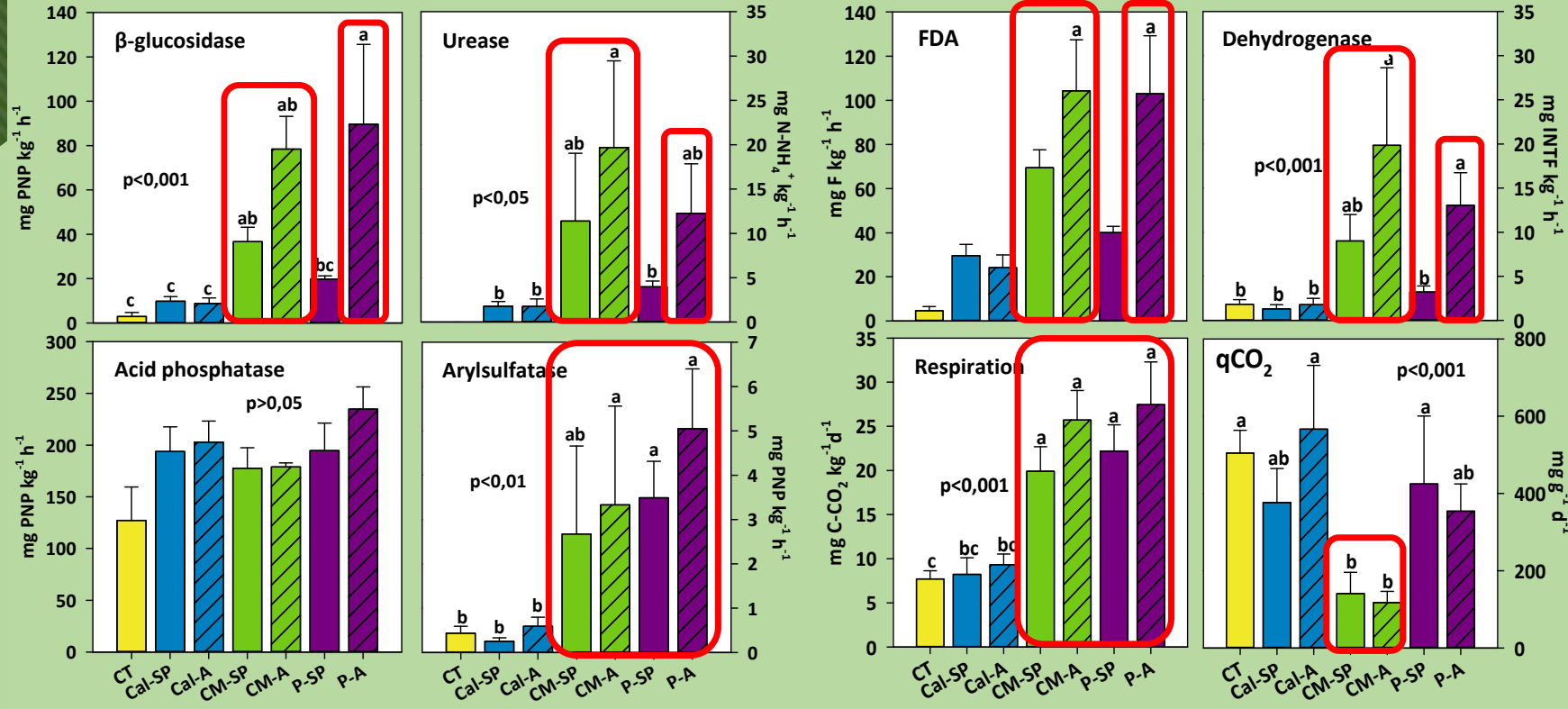
2 years after amendments addition



Case study 2 – Microbial indicators

Treatment	B _C mg kg ⁻¹	B _N mg kg ⁻¹
Cal-SP	24 c	0.3 c
Cal-A	20 c	0.2 c
CM-SP	167 a	10 a
CM-A	250 a	20 a
P-SP	29 bc	0.4 bc
P-A	91 ab	4.0 ab
CT	16 c	0.7 bc
T	***	**
PI	ns	ns
TxPI	***	***

Biomass



Functional activity: hydrolases (C, N & S cycle; catalytic potential (FDA); microbial community (dehydrogenase, respiration, qCO₂)

Case study 2 – Microbial indicators

CLPPs (community level physiological profiles): BIOLOG ECOPLATES



Average well color development (AWCD): overall functional activity of soil

Treatment	S	H'	AWCD
Cal-SP	6.0 ab	2.7 a	0.10 ab
Cal-A	7.0 ab	2.4 a	0.11 ab
CM-SP	14.8 a	3.6 a	0.38 a
CM-A	17.0 a	3.7 a	0.43 a
P-SP	8.0 ab	2.3 a	0.23 ab
P-A	12.3 a	3.2 a	0.23 ab
CT	1.2 b	0.4 b	0.01 b
T	***	***	
PI	ns	ns	
TxPI	**	**	

Functional diversity: S: species richness (number of substrates used); H': Shannon similarity index (uniformity in the distribution of the microbial community)

Case study 2 – Eco-toxicological tests

Plant growth test (direct) - *Zea mays* & *Lactuca sativa*

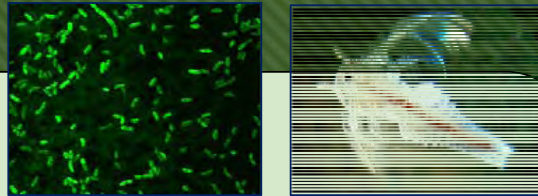


Treatment	<i>Z. mays</i>		<i>L. sativa</i>	
	LC ₅₀	EC ₅₀	LC ₅₀	EC ₅₀
Cal-SP	NT	85.75	58.19	20.02
Cal-A	NT	80.50	46.68	19.73
CM-SP	NT	NT	NT	18.31
CM-A	NT	NT	NT	NT
P-SP	NT	93.31	59.86	13.39
PCT-A	NT	91.70	NT	NT
CT	NT	70.85	36.68	4.95

LC₅₀ & EC₅₀: dose of contaminated soil (v/v) that provokes 50% emergence/growth inhibition, respectively. NT: non toxic

Case study 2 – Eco-toxicological tests

EC₅₀ (v/v)



Indirect assays:

- *Vibrio fischeri* (luminescence inhibition test)
- *Thamnocephalus platyurus* (mortality test)
- *Lepidium sativum* & *Lactuca sativa* (germination test)

Treatment	<i>V. fischeri</i>		<i>T. platyurus</i>
	t = 15 min	t = 30 min	
Cal-SP	62,52	57,97	78,83
Cal-A	60,76	59,77	73,53
CM-SP	62,55	61,15	93,82
CM-A	47,98	45,79	84,76
P-SP	54,01	49,51	82,28
P-A	62,29	60,86	78,78
CT	61,69	61,46	10,15

<i>L. sativum</i>			
Treatment	G (%)	R(%)	GI(%)
Cal-SP	87,8	51,8 d	45,4 c
Cal-A	84,7	51,9 d	43,9 c
CM-SP	91,8	64,0 c	58,7 b
CM-A	96,9	71,8 ab	69,6 a
P-SP	91,8	67,9 bc	62,3 ab
P-A	88,8	72,6 a	64,4 ab
CT	87,8	47,0 d	41,3 c
ANOVA	ns	***	***

<i>L. sativa</i>			
Treatment	G(%)	R(%)	GI(%)
Cal-SP	90	67,3 cd	60,5 bc
Cal-A	94	69,1 bcd	64,9 bc
CM-SP	99	92,9 abc	92 ab
CM-A	99	96,9 abc	95,9 a
P-SP	99	96,2 ab	95,2 a
P-A	98	105,8 a	103,6 a
CT	92	51,2 d	47,3 c
ANOVA	ns	**	**

Case study 2 – CONCLUSIONS

- ✓ Soil toxicity: high salinity, acidic pH and TEs solubility; amendments led to a **reduction of their potential associated risks**
- ✓ Supply of essential nutrients (pig slurry and compost) promoted a **stable vegetation cover**, that supposed an extra input of nutrients
- ✓ Both organic amendments in combination with *A. halimus* stimulated the soil microbial communities
- ✓ The combination of compost or pig slurry with *A. halimus*: effective strategy to **improve soil health** of TEs-contaminated soils under semi-arid conditions

Case study 3 - Restoration of a TEs contaminated agricultural soil



LOCATION OF EXPERIMENTAL SITE 3 (SG)

Characteristics	Soil SG
pH	7.6
OM (%)	3.9
Cu (mg kg ⁻¹)	28
Zn (mg kg ⁻¹)	632
Pb (mg kg ⁻¹)	651
Cd (mg kg ⁻¹)	<0.001
As (mg kg ⁻¹)	65

©2011 Google - Imágenes ©2011 DigitalGlobe, Cnes/Spot Image, GeoEye, European Space Imaging, Datos de mapa ©2011 Tele Atlas - Tér

Case study 3 - Restoration of a TEs contaminated agricultural soil

Fertilizer
1.3 t ha⁻¹

Compost
30 t ha⁻¹



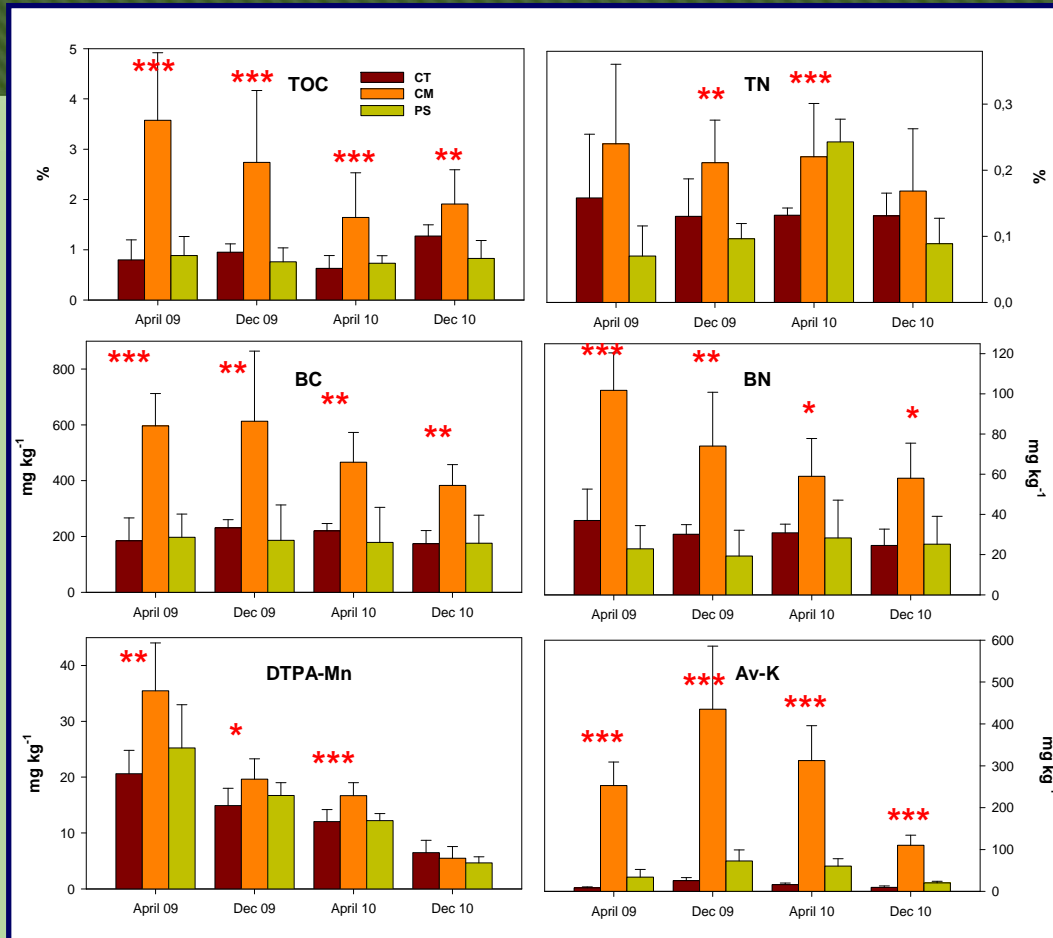
Case study 3 - Restoration of a TEs contaminated agricultural soil



~~*Atriplex confertifolia* L.~~

Bituminaria bituminosa L.

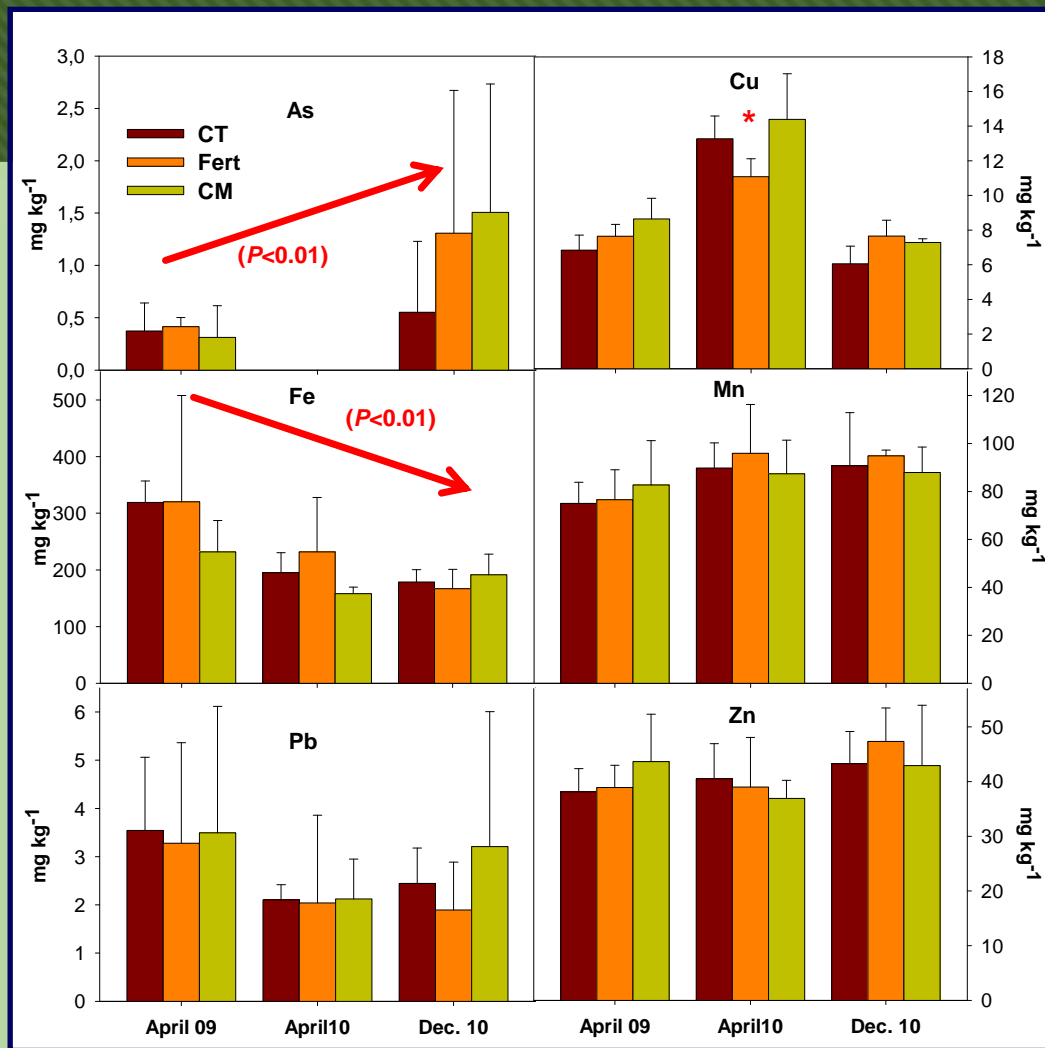
Case study 3 - SOILS: TREATMENT EFFECTS



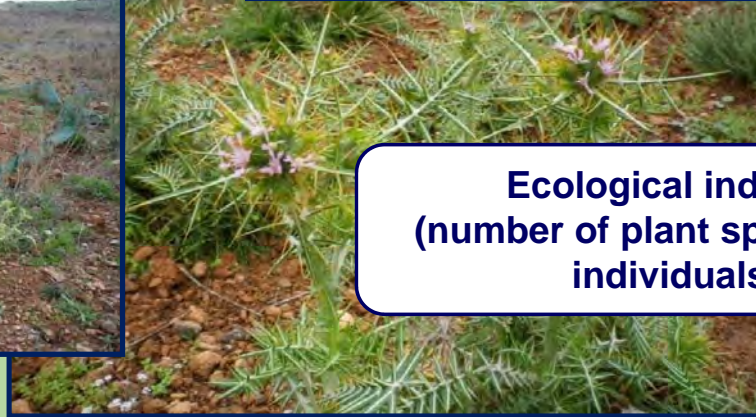
	Pb	Cd	As	C _{OT}	K	P
B _C	-0,347*	-0,307*	-0,458***	0,694***	0,771***	0,658***
B _N	-0,322*	-0,303*	-0,451***	0,758***	0,679***	0,661***

Pearson's Coefficient (r). *: p<0,05; **: p<0,01; ***: p<0,001

Case study 3 - PLANTS (*B. bituminosa*)



Case study 3 - PLANTS (spontaneous vegetation)



**Ecological indexes
(number of plant species and
individuals)**

Case study 3 - PLANTS (spontaneous vegetation)

Familia	Especie
Apiaceae	<i>Eryngium campestre</i>
Asteraceae	<i>Atractyllis cancellata</i>
	<i>Carduus bourgeanus</i>
	<i>Carthannus lanatus</i>
	<i>Crepis vesicaria subsp haenseleri</i>
	<i>Dittrichia viscosa</i>
	<i>Leontodon taraxacoides</i>
	<i>Phagnalon saxatile</i>
	<i>Reichardia tingitana</i>
Boraginaceae	<i>Cynoglossum creticum</i>
	<i>Echium creticum subsp. coincyantum</i>
	<i>Echium sabulicolum</i>
Brassicaceae	<i>Lobularia maritima</i>
Chenopodiaceae	<i>Atriplex halimus</i>
Cistaceae	<i>Fumana laevis</i>
Convolvulaceae	<i>Convolvulus althaeoides</i>
Crassulaceae	<i>Sedum sediforme</i>
Fabaceae	<i>Bituminaria bituminosa</i>
	<i>Coronilla scorpioides</i>
	<i>Scorpiurus sulcatus</i>
Frankeniaceae	<i>Frankenia corymbosa</i>
Myrsinaceae	<i>Anagallis arvensis</i>
Lamiaceae	<i>Lavandula multifida</i>
	<i>Thimus hyemalis</i>
Poaceae	<i>Phalaris brachystachys</i>
	<i>Stipa capensis</i>
Rubiaceae	<i>Galium verrucosum</i>
	<i>Valantia hispida</i>
Xanthorrhoeaceae	<i>Asphodelus fistulosus</i>
TOTAL	29

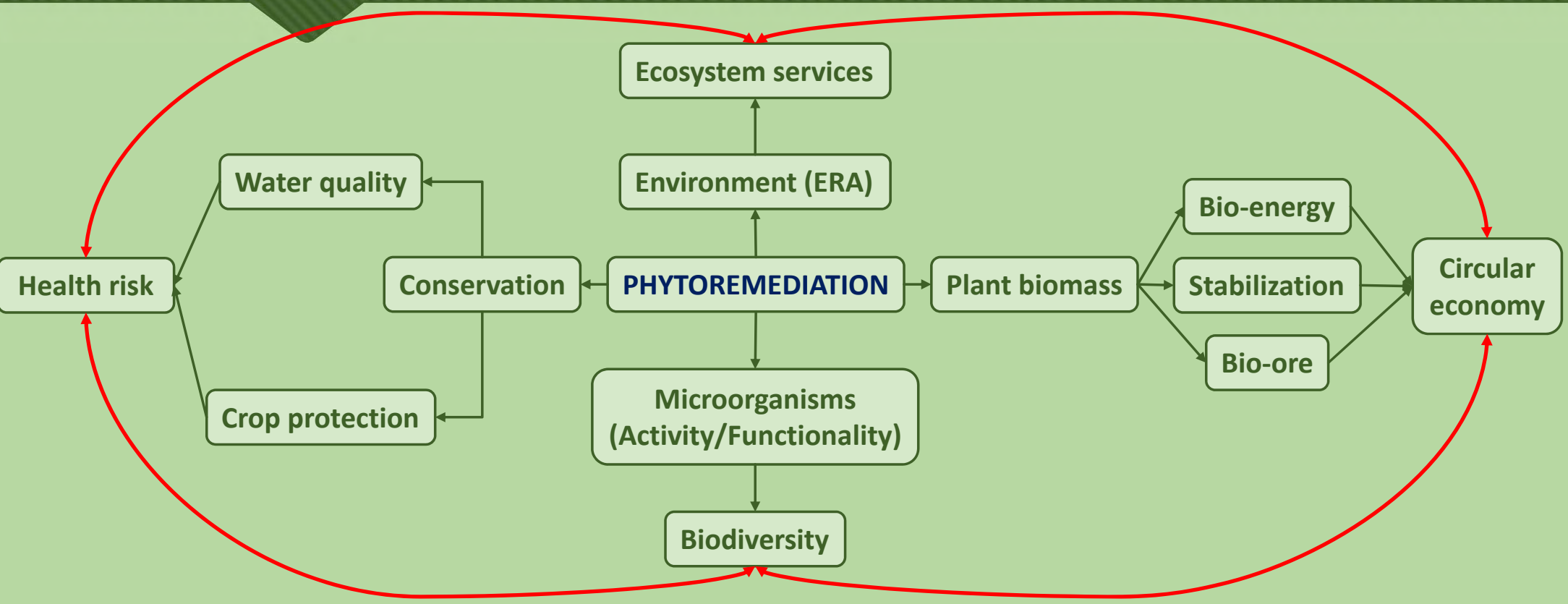
- ✓ 20.1% woody, 79.3% herbaceous
- ✓ No amendment effect on species distribution & abundance
- ✓ Compost > plant cover %



Case study 3 - **CONCLUSIONS**

- ✓ TEs present mostly in hardly-available forms; amendments application **did not alter extractability**
- ✓ Long-term supply of essential nutrients (compost): **development of the microbial community** and improved the plant growth (greater vegetation cover)
- ✓ Compost: **useful for phytostabilization** of moderately polluted soils, but combination with other species must be studied

Phytoremediation: Closing the loop



Acknowledgements



'Phytorec' team:

- Prof. M.P. Bernal
- Dr. T. Pardo
- Dr. E. Arco-Lázaro
- Ms. M.J. Álvarez-Robles
- Dr. D. J. Walker
- Dr. D. Martínez-Fernández
- Ms. D. Grippi

Collaborators:

- Prof. F. Sevilla (CEBAS-CSIC)
- Dr. X. Gómez (U. de León)
- Dr. S.Y. Huncce (Turkey)
- Ms. R. Chang (China)

THANK YOU!

School of Soil Biodiversity and Bioindication

*Biodiversity and bioindicators in monitoring
and management of contaminated soils*



GOBIERNO
DE ESPAÑA

MINISTERIO
DE CIENCIA, INNOVACIÓN
Y UNIVERSIDADES

